

APPLICATION
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TITLE: TISSUE-SPECIFIC AND TARGET RNA-SPECIFIC
RIBOZYMES

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**TISSUE-SPECIFIC AND
TARGET RNA-SPECIFIC RIBOZYMES**

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**TISSUE-SPECIFIC AND
TARGET RNA-SPECIFIC RIBOZYMES**

5 This application claims priority, under 35 U.S.C. § 119(e), of U.S. Provisional Patent Application No. 60/090,560 filed June 24, 1998, and of U.S. Provisional Patent Application No. 60/096,502 filed August 14, 1998, both of which are incorporated herein, by reference, in their entireties.

10 **1. FIELD OF THE INVENTION**

The present invention relates to multi-ribozymes and their use to target RNA in a tissue-specific or target-specific manner for the treatment of proliferative diseases, cancers and bacterial, parasitic and viral infections. More specifically, the present invention relates to the use of virions and viral vectors to package and deliver DNA encoding the multi-ribozymes to a host. The present invention relates to the use of liposomes and lipid-DNA complexes to deliver DNA encoding ribozymes to a host. Most specifically, the invention relates to the use of target specific virions to package and deliver DNA comprising a target specific promoter and encoding a ribozyme directed to target organism nucleic acids such as ribonucleic acids. The present invention further relates to a novel vector encoding a multi-ribozyme structure with enhanced 5' and 3' autocatalytically cleaving ribozymes.

2. BACKGROUND

2.1 RIBOZYMES

25 A ribozyme is a catalytic RNA molecule that cleaves RNA in a sequence specific manner. The use of ribozymes as potential gene regulators in mammalian cells and antiviral agents has been suggested, but subject to serious questions regarding technical feasibility. For example, there are differences regarding how ribozymes can be introduced to target cells or how they can be directed to the same subcellular compartments as their target 30 RNAs. Other questions concern the effects of target RNA secondary structure on ribozyme activity. The art has not been successful in definitively answering any of these questions.

Furthermore, because ribozymes are a form of antisense technology, the problems encountered in applying antisense technology to disease treatment are also encountered in the use of ribozyme technology (Haseloff, J., and W.L. Gerlach, 1988, *Nature* 334(6183):585-91; Sullivan, S. M., 1994, *J. Invest Dermatol.* 103(5 Supl):858-895). For 35 example, it has been shown that the expression of antisense RNA in transgenic mice did not invariably lead to a reduction in target RNA molecules, and when reduction in target RNA

molecules did occur, it was not predictably paralleled by a reduction in protein. Even when protein levels were reduced sometimes no biological effect was detected (Whitton, J. Lindsay "Antisense Treatment of Viral Infection" *Adv. in Virus Res.* Vol. 44, 1994).

The experience in the art suggests that it is also not clear whether ribozymes work best when free or when associated with only short non-specific flanking sequences, or when embedded in unrelated larger RNA molecule (Whitton, 1994, *supra*). At present, sufficient data are not available, either in *vitro* or in cell culture to allow systematic comparison of the transactivities of free ribozymes with their embedded equivalents.

There have been some studies that focus on the potential use of ribozyme technology in the treatment of cancer. In these studies, ribozymes have been directed against both c-fos and c-ras oncogenes in cell culture, and these showed some suppression of the malignant potential of transfected cells when transplanted into mice. Nevertheless, these ribozymes specifically target an oncogene.

There has been no suggestion in the literature that tissue-specific cancers or other tissue-specific disease can be treated by delivering to that tissue a ribozyme having a tissue-specific promoter, and that it is targeted to an RNA that is essential for cell survival. The invention provides such a ribozyme capable of treating tissue-specific cancers and other tissue-specific diseases.

20 2.2 PROMOTERS

Furthermore, endogenous delivery of a ribozyme under the control of a tissue-specific or other promoter can be complicated by "leakiness", where low levels of transcription occur in extraneous tissues. This could present a considerable therapeutic problem, depending upon the cellular target chosen. The present invention compensates for this problem by targeting a cellular target which is associated with high levels of product (such as, RNA polymerase I produces large amounts of cellular ribosomal RNA). Thus, in the event promoter leakiness occurs in unintended tissues, it is not likely that cell death would occur. This choice, therefore, provides a needed level of safety, and targeting of pol I would be applicable to many selected tissues using other promoters.

30

2.3 DELIVERY

Another common problem in gene therapy is the difficulty in delivering the ribozyme to the correct tissue. The present invention avoids this difficulty by targeting the ribozyme to non-cellular RNAs in cells to which ribozyme constructs can be efficiently delivered. IV liposome delivery will be effective for treatment of HBV hepatitis. Intravenous and/or extracorporeal treatment will effectively delivery construct to

erythrocytes for treatment of malarial infection. And topical (with or without iv) administration will effectively deliver ribozyme construct to cervical epithelium in dysplastic/precancerous/cancerous HPV 16 cervical lesions. This latter example is of extreme importance for treatment of dysplastic/carcinoma *in situ* lesions diagnosed via 5 abnormal Pap smears. A second advantageous facet of the non-cellular target ribozymes of the present invention is that even if promoter leakiness and/or extraneous delivery and/or expression of the ribozyme occurs in unintended cells, the ribozymes should not cleave the cellular RNAs.

10 **2.4 ANTIMICROBIAL AGENTS**

Infectious diseases sicken or kill millions of people each year. Numerous antimicrobial therapies have been designed to target one or several infectious agents. These therapies show varying degrees of success in eradicating infection. However, the failure of many of these therapies to target specific infectious agents has lead to overuse or 15 inappropriate use of the therapies, which in turn has lead to the development of drug resistant microbes. The development of drug resistance in many infectious agents has reduced the efficacy and increased the risk of using the traditional antimicrobial therapies.

While ribozymes have been known and studied for several years, they have not been used in the treatment of bacterial infections. There are many reasons for this. A key 20 technical concern in the use of ribozymes as antimicrobial agents is that the ribozyme must be taken up and expressed by the targeted microbe so that the ribozyme(s) can cleave the targeted RNA(s) inside the microorganism. A second important concern is the tight coupling of transcription and translation in microorganisms which can complicate efficient cleavage of targeted bacterial RNAs. These concerns are addressed by the present 25 invention.

3. SUMMARY OF THE INVENTION

The present invention relates to multi-ribozymes and their use to target RNA in a tissue-specific or pathogen-specific manner for the treatment of cancers and proliferative 30 disease and bacterial, parasitic and viral infections. The present invention further relates to multi-ribozymes that target pathogens.

The present invention relates to multi-ribozyme(s) which contain two separable functional regions including a "catalytic core" which cleaves the target RNA or RNAs, and flanking regions which include cis-acting autocatalytically acting ribozymes and a target 35 RNA-specific binding site. The catalytic core contains one or more ribozymes known as trans-acting ribozymes. The flanking regions are located nearby or adjacent to the catalytic

core, and contain ribozymes known as autocatalytically cleaving ribozymes. A catalytic core in combination with one or more flanking region(s) as used herein is referred to as a ribozyme "cassette" or "triple ribozyme" (when three ribozymes comprise a multi-ribozyme.). In several embodiments, a multi-ribozyme comprises one or more ribozyme 5 cassette. By nucleic acid complementarity, the binding site directs the multi-ribozyme core to cleave a specific site on the target RNA molecule.

The invention also relates to multi-ribozyme structures containing modifications which enhance stability and protect against degradation, *e.g.*, RNA hairpin loops. Examples of such modifications include those which protect against degradation by endonucleases 10 such as modifications to the structure of the nucleotides and stabilizing hairpin loops in or near the ribozyme cassette.

The present invention relates to novel vectors encoding multi-ribozymes of the invention. The invention further relates to the use biologic delivery systems such as virions and viral vectors to package and deliver the DNA encoding the multi-ribozymes for the 15 present invention. The invention encompasses the use of abiologic delivery systems such as liposomes and liposome-DNA or lipid-DNA complexes to deliver DNA encoding ribozymes to a host.

The invention provides tissue-specific and target RNA-specific (such as pathogen-specific) ribozymes. The ribozymes can be targeted specifically to neoplastic cells or viral 20 infected cells in order to target RNA encoding essential gene products, and thus eliminate the neoplastic or viral infected cell. The ribozymes of present invention provides multi-ribozymes which are designed as combinations of autocatalytic and trans-acting ribozymes. The present invention further provides autocatalytically cleaving ribozymes with enhanced activity. In accordance with the present invention the multiple trans-acting 25 ribozymes may be targeted to the same site on the same RNA, different sites on the same RNA, or different RNAs.

The present invention further provides for regulating the cellular distribution of the transacting ribozymes. The present invention encompasses the use of combinations of slow cleaving and enhanced cleaving autocatalytically cleaving ribozymes to regulate the nuclear 30 and cytoplasmic accumulation and distribution of the trans-acting ribozymes.

The invention additionally provides nucleic acids and cassettes which encode the ribozymes of the invention. These nucleic acids can be used to express the ribozymes of the invention at a selected site. In several embodiments of the invention, the nucleic acids or expression cassettes encode a tissue-specific or target-specific promoter operably linked to a 35 nucleic acid encoding a multi-ribozyme of the invention. In one embodiment of the invention, the nucleic acids or expression cassettes of the invention comprise a tissue-

specific promoter upstream from a sequence encoding a 5' autocatalytically cleaving ribozyme sequence, a catalytic ribozyme comprising a target RNA-specific binding site and a 3' autocatalytically cleaving ribozyme sequence. In another embodiment of the invention, the nucleic acids and expression cassettes of the invention comprise a tissue-specific 5 promoter upstream from a sequence encoding a 5' autocatalytically cleaving ribozyme sequence, a catalytic ribozyme comprising one or more target RNA-specific trans-acting ribozymes and a 3' autocatalytically cleaving ribozyme sequence.

In accordance with the present invention, the expression cassettes may be engineered to express two or more multi-ribozymes containing transacting ribozymes which 10 act on the same or different targets. The expression cassettes may also be engineered to express one, two or more multi-ribozymes containing 5' and 3' autocatalytically cleaving ribozymes with either slow or enhanced cleavage activity.

In a preferred embodiment, the invention provides nucleic acids and expression cassettes which encode multi-ribozymes with altered cleavage sites, so that the 5' and/or 3' 15 autocatalytically cleaving ribozymes have enhanced activity, resulting in the more effective and efficient release of the targeted internal ribozymes. In an additional preferred embodiment, the invention provides nucleic acids which encode multi-ribozymes with one or more trans-acting ribozymes, resulting in the more effective and efficient targeting of RNA-target(s). In other embodiments, the invention provides for nucleic acids that encode 20 one or more ribozyme cassettes each containing a) a 5' autocatalytically cleaving ribozyme sequence and/or a 3' autocatalytically cleaving ribozyme; and b) catalytic ribozyme(s) comprising one or more target RNA-specific trans-acting ribozymes. In another embodiment, the expression cassettes encode autocatalytically cleaving ribozyme combinations of slow and enhanced cleavage activities thus resulting in a distribution of 25 liberated trans-acting ribozymes between the nucleus and cytoplasm. In yet another embodiment, the expression cassette encodes enhanced autocatalytically cleaving ribozymes resulting in an increase accumulation of the liberated trans-acting ribozymes in the nucleus.

In another preferred embodiment, the present invention relates to a microbiocidal 30 agent directed against any cellular, viral, bacterial, fungal, or other single or multicellular organism from any known taxonomic family, genus, or species, and from previously unknown, or uncharacterized organism. In yet another preferred embodiment, the present invention relates to a multi-ribozyme comprising a trans-acting ribozyme which targets any cellular, viral, bacterial, fungal, or other single or multicellular organism from any known 35 taxonomic family, genus, or species, and from previously unknown, or uncharacterized organism. The present composition of matter has resulted from the development of a new

delivery system that provides a series of ribozymes directed against fundamental and essential cellular processes specific to a targeted microorganism through an inactivated, altered, virus (virion), or biologic delivery vehicles, capable of delivering a nucleic acid containing the ribozyme(s) into the targeted microorganism. The targeted microorganisms 5 may be any virus, nonvirus, bacterium, or lower eukaryotes such as fungi, yeast, parasites, protozoa, or other eukaryotes that may be considered normal flora or pathogens of humans, animals, fish, plants or other forms of life.

The present invention also relates to a Multi-ribozyme ribozyme. The present Multi-ribozyme ribozyme is uniquely suited as an antimicrobial therapeutic in that upon 10 nucleic acid hybridization with the target RNA transcript, the ribozyme-RNA complex achieves a catalytic form that acts as a nuclease to cleave the targeted RNAs. Thus, cleavage deprives the invading microorganism of essential cellular processes which then kill or render it less fit.

This approach offers new and unprecedented advances for antimicrobial 15 therapeutics: 1) the preparation bypasses any de novo built-in drug resistance, which sophisticated microbes will be expected to have or develop, 2) cells are generally not capable of counteracting ribozymes delivered into them, 3) microbes have several broad RNA targets that can be attacked in simultaneously with probable synergy, 4) the custom design of the present delivery vehicle can be readily tailored to different families of 20 organisms, 5) the modified delivery vehicle is a non-replicating, artificial construct easy to assemble and manufacture, 6) the preparation can be applied topically or it can be delivered via injection, inhalation, or ingestion, 7) the preparation can be lyophilized and thus confer stability to the antimicrobial therapeutic, 8) the inhalant, ingested or topical form of the antimicrobial therapeutic reduces the immunogenicity of the multi-ribozyme preparations as 25 opposed to its parenteral use, and 9) animal test systems exist that enable the evaluation of the multi-ribozyme in a measured, incremental fashion to quickly determine the efficacy of the antimicrobial therapeutic agent. Therefore, the combination of the present unique delivery approach and an aggressive mechanism for depriving the microbial cells of essential gene products can achieve the timely defeat of microbes within the host.

30 The targets of the antimicrobial multi-ribozyme therapeutic described herein are the RNAs of invading or normal flora microorganisms. The invention provides the delivery of a series of ribozymes directed towards essential, housekeeping, or virulence genes of one or a series of candidate microorganisms. A ribozyme is uniquely suited as the active component of the present antimicrobial therapeutic in that it is a catalytic RNA molecule 35 that cleaves RNA in a sequence specific manner. Therefore, the catalytically active component of a multi-ribozyme contains ribozymes that have been designed to inactivate

RNA coding for components of the microbial cell. Inactivation of essential proteins and virulence determinants render the invading microbes inactive or slow their growth, while at the same time, the essential processes of the host are not affected.

At the molecular genetic level, the coding sequence for a ribozyme or the multi-
5 ribozyme may be placed under the control of one or more of the following genetic elements:
a naturally occurring strong, intermediate or weak constitutively expressed or regulated
promoter from the targeted microorganism, or an artificially contrived constitutively
expressed or regulated promoter containing either a strong, intermediate or weak consensus
sequence that delivers desired levels of ribozyme expression. This genetic information is
10 delivered into the microbe by either a biologic (e.g., a modified virus) or abiologic delivery
vehicle.

In one embodiment of the present invention, when a biologic vehicle is used, the
nucleic acids encoding the multi-ribozymes are unique in that they contains sufficient
genetic information for expression of the ribozyme(s) and such genetic information
15 necessary and sufficient for its assembly and delivery to the targeted microorganism, but
does not include nucleic acids native to the virus. Thus, the virion can serve as a molecular
vehicle that delivers the inactivating ribozyme(s). Alternatively, an abiologic delivery
system (e.g., liposomes) can be used to package nucleic acid carrying the genetic elements
necessary and sufficient for the proper expression of the ribozyme(s).

20 In yet another embodiment, the present invention relates to a novel vector encoding
multi-ribozyme structures. The present invention also relates to a novel vector encoding
multi-ribozyme structures with enhanced 5' and/or 3' autocatalytically cleaving ribozymes.
The novel vectors of the present invention encode unique 5' and 3' autocatalytically cleaving
activity, so that the internally encoded ribozymes are rapidly and effectively released. The
25 novel vectors of the present invention may be used to engineer a wide variety of ribozymes
including, but not limited to, tissue and/or promoter specific ribozymes, anti-microbial
ribozymes, anti-viral ribozymes, anticancer ribozymes, anti-proliferative ribozymes, anti-
tumor ribozymes.

The present invention further encompasses the use of the multi-ribozymes of the
30 present invention for the treatment of disease, viral infection, parasitic infection, and
microbial infection. The present invention further encompasses the use of the multi-
ribozymes of the present invention for the treatment of proliferative disease such as
neoplasms, malignancies, and other cancers. The present invention further relates to a
method of treating a subject having a proliferative disease of a specific tissue by inhibiting
35 cell proliferation in the tissue, comprising administering to the subject a multi-ribozyme
wherein the target-specific promoter binding sequence is specific for the diseased tissue,

whereby the ribozyme encoded by the nucleic acid is expressed, ribosomal RNA production in the tissue is inhibited, cell proliferation is inhibited, and the proliferative disease is treated or ameliorated.

The present invention further relates to a method of treating a subject having a proliferative disease of a specific tissue by inhibiting cell proliferation in the tissue, comprising administering to the subject a multi-ribozyme wherein the tissue-specific promoter binding sequence is specific for the diseased tissue, whereby the ribozyme(s) encoded by the nucleic acid is expressed, ribosomal RNA production in the tissue is inhibited, cell proliferation is inhibited, and the proliferative disease is treated or ameliorated.

The present invention encompasses the multi-ribozymes of the present invention in pharmaceutical formulations.

The present invention encompasses the use of the multi-ribozymes of the present invention for research and screening purposes. In one embodiment of the present invention, the multi-ribozymes may be used to screen for viral or microbial gene products to be targeted in order to effectively inhibit the life cycle of the virus or microbial agent. In another embodiment of the invention, a library of sequences (e.g., a ribozyme library) is used to screen for trans-acting ribozymes that may be directed to a particular target RNA or an unknown RNA target.

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4. BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1A shows a schematic of DNA encoding the ribozyme used in the molecular sequence of events in ribozyme maturation and action.

Figure 1B shows the primary RNA transcript. Autocatalytic cleavage takes place upon completion of transcription.

Figure 1C shows the release of the trans acting ribozyme. As a direct result of cleavage of the two cis-acting ribozymes, the internal ribozyme containing a reverse and complementary sequence to the mRNA target is released.

Figure 1D shows the sequence specific hybridization of the ribozyme. The internal or trans-acting ribozymes comprise two trans-acting ribozymes linked by a short nucleotide "spacer". Each of the two trans-acting ribozymes contain a sequence that is reverse and complementary to the targeted message. The ribozyme is synthesized at a concentration sufficient to locate and hybridize to all or substantially all targeted transcripts.

Figure 1E shows the trans-catalytic cleavage. Upon hybridization of the internal ribozyme to the targeted mRNA transcript, the internal ribozyme achieves a catalytic

topology and cleaves the targeted message. Upon cleavage the trans acting ribozyme is released and its activity and function are recycled.

Figure 2 Diagram and nucleotide sequence of the pClip ribozyme cassette.

Figure 3 Diagram and nucleotide sequence of the pChop ribozyme cassette.

5 Figure 4 Schematic diagram of the pSnip ribozyme cassette. pSnip includes sequences of the pClip triple ribozyme cassette, catalytic core targeted ribozymes comprising two linked trans-acting ribozymes, and sequences from the pChop triple ribozyme cassette

10 Figure 5 Nucleotide sequence of pSnip, shown without the double trans-acting ribozyme inserts.

15 Figure 6 Schematic diagram of various ribozyme constructs which were synthesized for testing *in vitro*. V denotes vector flanking sequences, 5'AC denotes the 5' autocatalytic trans-acting ribozyme which was modified so that self-liberation could not occur. ITRz denotes the internal trans-acting ribozymes also modified so that self-liberation could not occur.

20 Figure 7 *In vitro* Analysis of Cleavage Activity of Multi-Ribozymes. 50nM of the each of the constructs 1 through 6 as shown in Figure 6 were incubated for 37°C for 0.05 or 2 hours with 50nM of target RNA. In this example, the transacting ribozymes of the constructs were targeted to multi-catalytic proteinase component C9. Following incubation, samples were denatured and separated on a polyacrylamide gel. The upper band represents the uncleaved target RNA whereas the lower bands represent the cleaved products. The data demonstrate that a ribozyme attached at either the 5' (lane 2) or 3' (lane 4) end of a transacting ribozyme does not diminish the catalytic activity of the ribozyme.

25 Figure 8 RT/PCT Analysis of TRz Expression and Self-Liberation in B2-X Clones. Cells were stably transfected with a B2-targeted PCLIP multi-ribozyme. Figure 8A shows cytoplasmic RNA results, while Figure 8B shows nuclear RNA results. These results demonstrate that there is a distribution of liberated transacting ribozymes between the nucleus and the cytoplasm.

30 Figure 9 Strategy of screening a ribozyme library to identify trans-acting ribozymes and/or targets.

Figure 10 Sequence of a modified pChop cassette (UPCM2).

Figure 11 *In vitro* substrate cleavage results of ribozymes targeted to NfkB p50 subunit.

35 Figure 12 Effect of Ribozyme (RZ1024, targeted to NfkB p50 subunit) on TNF α - induced IL-6 production.

5. DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to multi-ribozymes and their use to target RNA in a tissue-specific or target-specific manner for the treatment of disorders and disease related to cellular proliferation, cancers and bacterial, parasitic, or viral infections. The multi-ribozymes of the present invention may be engineered to target one or more specific RNAs contained in a specific cell or tissue in the host. The multi-ribozymes of the present invention may also be engineered to target one or more specific RNAs encoded by a specific pathogen, virus or microbial agent.

The multi-ribozyme(s) of the invention contain two separable functional regions including a "catalytic core" which cleaves the target RNA or RNAs, and flanking regions which include a target RNA-specific binding site. The catalytic core contains one or more ribozymes known as trans-acting ribozymes. The flanking regions are located nearby or adjacent to the catalytic core, and contain ribozymes known as autocatalytically cleaving ribozyme sequences or autocatalytic ribozymes. A catalytic core in combination with one or more flanking region(s) as used herein is referred to as a ribozyme "cassette" or "triple ribozyme". By nucleic acid complementarity, the binding site directs the multi-ribozyme core to cleave a specific site on the target RNA molecule.

In accordance with the present invention the multi-ribozymes may comprise a 5' autocatalytically cleaving ribozyme sequence, a catalytic ribozyme comprising a target RNA-specific binding site and a 3' autocatalytically cleaving ribozyme. In accordance with the invention, the multi-ribozymes may comprise a) a trans-acting ribozyme flanked by 5' and 3' autocatalytically cleaving ribozymes or flanked by enhanced 5' and 3' autocatalytically cleaving ribozymes; b) a transacting ribozyme flanked by either a 5' or 3' autocatalytically cleaving ribozyme or flanked by an enhanced 5' or 3' autocatalytically cleaving ribozyme; or c) multiple trans-acting ribozymes flanked by one or both 5' and/or 3' autocatalytically cleaving ribozymes or flanked by enhanced 5' and/or 3' autocatalytically cleaving ribozymes.

In accordance with the present invention, the multi-ribozymes may be designed to release two or more trans-acting ribozymes. Such trans-acting ribozymes may be targeted to the same site on the same RNA, different sites on the same RNA, or different RNAs. In an additional embodiment, the ribozyme cassettes are linked in a series of one or more. In another embodiment, the linked ribozyme cassettes are non-identical.

In particular, the multi-ribozymes of the present invention are designed to have improved properties to enhance their efficacy in cleaving a target RNA. The multi-ribozymes of the present invention comprise a) one or more transacting ribozymes and b) 5' and 3' flanking autocatalytically cleaving ribozymes or only the 5' or the 3'

flanking autocatalytically cleaving ribozyme. The flanking autocatalytically cleaving ribozymes act to liberate the transacting ribozyme. The autocatalytically cleaving ribozymes of the present invention may have slow cleavage activity or enhanced cleavage activity. The combination of slow cleaving autocatalytic ribozymes followed by enhanced 5 cleaving autocatalytic ribozymes results in the distribution of transacting ribozymes between the nucleus and the cytoplasm. The use of enhanced cleaving autocatalytic ribozymes results in primarily a nuclear accumulation of the trans-acting ribozymes and the use of slow cleaving ribozymes results in primarily a cytoplasmic accumulation of the trans-acting ribozymes.

10 In one preferred embodiment, ribozyme cassette(s) lead to distribution of the liberated trans-acting ribozyme(s) between the nucleus and cytoplasm. In another preferred embodiment, the ribozyme cassettes lead to distribution of the liberated trans-acting ribozyme(s) with increased distributed to the nucleus.

15 Use of a "spacer" for linking the ribozymes together is provided by the invention. In one specific embodiment, but not by way of limitation, the ribozymes and/or ribozyme cassettes are linked together by a short "spacer" of 4-5 nucleotides.

20 The invention also provides multi-ribozymes containing modifications which enhance stability and protect against degradation. Examples of such modifications include those which protect against degradation by endonucleases such as modifications to the structure of the nucleotides and stabilizing hairpin loops in or near the ribozyme cassette. In one embodiment, the of the present invention one or more ribozymes are stabilized by a 3' hairpin loops.

25 The present invention relates to nucleic acids encoding the tissue-specific or target RNA-specific ribozymes of the present invention. In one embodiment of the invention, the nucleic acids of the present invention may comprise a tissue-specific promoter operably linked to a multi-ribozyme of the invention. In one embodiment of the invention, the nucleic acids of the present invention may comprise a tissue-specific promoter upstream from a sequence encoding a 5' autocatalytically cleaving ribozyme, a catalytic ribozyme comprising a target RNA-specific binding site and a 3' autocatalytically cleaving ribozyme 30 sequence. In another embodiment of the invention, the nucleic acids may comprise a tissue-specific promoter upstream from a sequence encoding a 5' autocatalytically cleaving ribozyme, an internal targeted catalytic core containing two or more target RNA-specific trans-acting ribozymes and a 3' autocatalytically cleaving ribozyme sequence.

35 In a preferred embodiment, the nucleic acid encodes the modified 5' and/or 3' autocatalytically cleaving ribozymes of the present invention which have enhanced cleaving

activity which results in the enhanced and more effective release of the internal catalytic ribozyme(s).

The present invention further relates to the use of a wide variety of vehicles to deliver the multi-ribozymes to a target, including biologic vehicles such as virions and viral vectors to package and deliver the DNA encoding the multi-ribozymes; non-viral expression vectors and non-biologic (abiologic) vehicles, including liposomes and liposome-DNA and lipid-DNA complexes to deliver and target the DNA encoding the multi-ribozymes to the host.

In another embodiment, the invention relates to nucleic acids which encode multi-ribozymes which are targeted by their delivery vehicle.

In accordance with the invention, the host to which the multiple-ribozymes are delivered may be cells in culture, tissues in culture, plants, animal models, animals, mammals or humans.

The present invention relates to pharmaceutical compositions comprising the multi-ribozymes of the present invention and their delivery vehicles. The multi-ribozymes of the present invention may be engineered for the treatment of a wide variety of disorders and diseases related to expression of a particular gene or genes, cellular overproliferation, hereditary disorders, cancers, tumors, viral infections, bacterial or parasitic infections.

In another embodiment of the present invention, the multi-ribozymes of the present invention may also be used for *in vitro* screening purposes, e.g., to identify a gene product involved in cellular overproliferation or to identify a gene product critical for the life cycle of a viral or microbe.

5.1. RIBOZYMES

The present invention provides tissue-specific and target RNA-specific ribozymes. These ribozymes can be used to destroy tissue-specific neoplasms, cancers, or proliferative disorders and to treat viral, bacterial or parasitic infections, among other uses. The multi-ribozymes of the present invention comprise one or more ribozymes or ribozyme cassettes.

In accordance with the present invention, the multi-ribozyme may consist of one or more ribozyme cassettes. Each cassette in turn may consist of a catalytic core and one or more flanking sequences. In one embodiment, the ribozyme cassette may consist of a 5' autocatalytically cleaving ribozyme sequence, a catalytic ribozyme comprising a target RNA-specific binding site and a 3' autocatalytically cleaving ribozyme. In another embodiment, the ribozyme cassette may consist of a 5' autocatalytically cleaving ribozyme sequence, a catalytic core ribozyme comprising a trans-acting ribozyme and a 3' autocatalytically cleaving ribozyme.

In a preferred embodiment, the multi-ribozyme comprises an enhanced 5' and 3' autocatalytically cleaving ribozyme sequence. In another preferred embodiment of the invention, the multi-ribozyme comprises the ribozyme cassette as shown in Figure 2. In another preferred embodiment, the multi-ribozyme comprises the ribozyme cassette as 5 shown in Figure 3. In a most preferred embodiment, the multi-ribozyme comprises the ribozyme cassette as shown in Figure 4. In another embodiment, the ribozymes are in a cassette. In yet another embodiment, the multi-ribozymes contain two, three, four or more internal trans-acting ribozymes. In a preferred embodiment, the multi-ribozymes of the present invention include, but are no limited to triple ribozyme cassettes (as described 10 herein, such as a cassette composed of one or more trans-acting ribozymes linked to a 3' and 5' flanking autocatalytically cleaving ribozymes). In another embodiment, multi-ribozymes include but are not limited to one or more triple ribozyme cassettes linked together. In an additional embodiment, the multi-ribozyme comprises a series of one or more ribozyme cassettes containing one or more internal trans-acting ribozymes or any combination 15 thereof. In one specific embodiment, the multi-ribozyme comprises two ribozyme cassettes, each with two trans-acting ribozymes in the catalytic core (Double ITRz) as shown in Figure 4.

The invention provides ribozymes that have the unique characteristic of being both target RNA-specific in their catalytic action, and subject to tissue-specific expression.

20 Examples of the enhanced 5' and 3' autocatalytically cleaving ribozymes that are expressed with the catalytic ribozyme of the invention are also shown in Figure 3. Other examples of nucleotides encoding 5' and 3' autocatalytically cleaving ribozymes and vectors for the multi-ribozymes of the present invention are described in WO 97/17433, incorporated herein by reference in its entirety. As further described below, these autocatalytically 25 cleaving ribozymes are important for the expression of the catalytic ribozyme, because they cleave off the ribozyme transcript as soon as they are transcribed to produce a catalytic ribozyme having minimal extraneous 5' or 3' sequences. Further, the enhanced autocatalytically cleaving ribozymes are important for the expression of the trans-acting catalytic ribozyme, influencing the cellular distribution of the transacting ribozymes and 30 since they cleave the ribozyme transcript as soon as they are transcribed to produce a catalytic trans-acting ribozyme having minimal extraneous 5' or 3' sequences.

Thus, the target-specific binding site and the catalytic sequence that comprise the catalytic ribozyme are in the correct configuration to bind and cleave the target RNA. The extraneous sequences in the exemplified construct are the result of the cloning procedure. It 35 is understood that with the selection of an alternative cloning scheme some or all of these extraneous nucleotides can be eliminated.

The Applicants have demonstrated that liberating the internal ribozyme is five to ten fold more active with the cleaving auto-catalytic ribozymes than when the same ribozyme is embedded within other flanking sequences. Applicants have further demonstrated that liberation of internal trans-acting ribozymes from a combination of slow cleaving and 5 enhanced cleaving auto-catalytic ribozymes results in a distribution of the trans-acting ribozyme between the nucleus and cytoplasm.

5.1.1. RIBOZYME-ENCODING NUCLEIC ACIDS

The invention also provides nucleic acids which encode the ribozymes of the 10 invention. The invention provides nucleic acids which encode ribozyme(s) which are tissue-specific or target RNA-specific. The invention also provides nucleic acids which encode ribozyme(s) operably linked to a tissue-specific, or target-specific (*i.e.*, pathogen-specific) promoter. These nucleic acids can be used to express the ribozymes of the invention at the selected site. For example, the site can be tissue-specific in the case of 15 treating tissue-specific cancers or disease, or it can be target-specific in the case of ribozymes that prevent replication of infectious agents to treat infection (*e.g.* hepatitis, herpes, malaria, tuberculosis, etc.).

In several embodiments, nucleic acids of the invention encode a catalytic multi-ribozyme that contains two separable functional regions including a) a catalytic core 20 sequence which cleaves the target RNA, and b) flanking regions which include cis-acting autocatalytically cleaving ribozyme(s). The length of flanking sequences have implications not only for specificity, but also for the cleavage efficiency of the individual ribozyme molecules. In the present catalytic ribozyme, the flanking sequences are highly specific for the target RNA, yet allow ready dissociation from the target RNA once cleavage occurs. 25 This permits cycling of the ribozyme (with an expected Kcal of about 1 cleavage per minute) and reduces the amount of ribozyme required to be effective. A range of binding/dissociation values from 16-21 Kcal is expected to be effective.

In accordance with the invention, a pathogen-specific promoter or tissue-specific promoter may be operably linked to any sequence encoding a multi-ribozyme or expression 30 cassette of the invention. In several embodiment, the nucleic acids of the invention comprise a tissue-specific promoter operably linked to a sequence encoding a multi-ribozyme. The tissue-specific promoter in the ribozyme-producing construct results in tissue-specific expression of the ribozyme(s) in tissue(s) that actively transcribe RNA from the selected promoter. Thus, only the target RNA in tissue that utilize the promoter will be 35 cleaved by the ribozyme(s).

In other embodiments, the nucleic acids and expression cassettes of the invention comprise a pathogen-specific promoter operably linked to a sequence encoding a multi-ribozyme. For example, a pathogen-specific promoter or tissue-specific promoter may be operably linked to a nucleotide sequence encoding a) a catalytic core ribozyme sequence 5 which cleaves the target RNA of the pathogen, and b) 3' and/or 5' flanking regions which include cis-acting autocatalytically cleaving ribozyme(s).

In accordance with the present invention, a multi-ribozyme may be engineered to express two or more ribozyme cassettes containing trans-acting ribozymes which act on the same or different targets. In an additional embodiment, the invention provides for nucleic 10 acid that encode one or more ribozyme cassettes each cassette containing a) a 5' autocatalytically cleaving ribozyme sequence and/or a 3' autocatalytically cleaving ribozyme sequence; and b) one or more catalytic ribozymes comprising one or more target RNA-specific trans-acting ribozymes. In a preferred embodiment, the invention provides nucleic acids which encode multi-ribozymes with multiple trans-acting ribozymes, resulting 15 in the more effective and efficient cleavage of target RNA.

In other embodiments, the invention provides nucleic acids and expression cassettes which encode multi-ribozymes with altered cleavage sites, so that the 5' and/or 3' autocatalytically cleaving ribozymes have enhanced activity, resulting in the more effective and efficient release of the internal trans-acting ribozymes. Thus, the expression cassettes 20 may also be engineered to express two or more multi-ribozymes containing 5' and/or 3' autocatalytically cleaving ribozymes with either slow or enhanced cleavage activities. In one embodiment, expression cassette encodes a combination of autocatalytically cleaving ribozymes with slow and enhanced cleavage activities, resulting in a distribution of liberated trans-acting ribozymes between the nucleus and cytoplasm of a eukaryotic cell. In 25 another embodiment, the expression cassette encodes enhanced autocatalytically cleaving ribozymes resulting in an increase accumulation of the liberated trans-acting ribozymes in the nucleus.

The nucleic acid, wherein at least one triple ribozyme is targeted to the *rpoA* transcript of the pathogen is provided. The nucleic acid, wherein at least one triple ribozyme 30 is targeted to the *secA* transcript of the pathogen is provided. The nucleic acid, wherein at least one triple ribozyme is directed to the *dnaG* transcript of the pathogen is provided. The nucleic acid, wherein at least one triple ribozyme is directed to the *ftsZ* transcript of the pathogen is provided. A ribozyme-encoding nucleic acid can encode all or some of the above triple ribozymes. The triple ribozymes and ribozyme cassettes can all be under the 35 control of a single promoter.

Many examples of the nucleic acid encoding the trans-acting ribozyme of the triple ribozyme are described herein and in the Sequence Listing (e.g., SEQ ID NO:8 -17, 43-48) and Examples Section herein.

5 **5.1.2 PROMOTER SELECTION**

Promoter selection is accomplished using techniques that are available in the art. As used herein, regulatory elements include but are not limited to, inducible and non-inducible promoters, enhancers, operators and other elements known to those skilled in the art that drive and regulate expression. Specifically, the invention provides inducible promoters 10 which have increased transcriptional control and high expression levels. The promoter can be a naturally occurring strong, intermediate or weak constitutively expressed or regulated promoter from the targeted microorganism, or an artificially contrived constitutively expressed or regulated promoter containing either a strong, intermediate or weak consensus sequence that delivers desired levels of ribozymes in the targeted microbe. For example, a 15 method is described in the Examples that permits the selection of both controlled and uncontrolled promoters, as well as consensus promoters that can be design for application in the present multi-ribozyme.

Promoters specific for the target (e.g., a specific pathogen, genus, etc.) in question can be selected by screening genomic sequences for the ability to activate a promoterless 20 reporter gene. The promoterless reporter gene is based on the strategy developed for use with plasmid pMC1871 (Casadaban et al., 1983, Meth. Enzymol. 100:293). For non-viral pathogens, plasmid capable of stable replication and maintenance in the microorganism under study is modified by standard molecular biology techniques to carry the coding region of a reporter gene (Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., 25 Cold Spring Harbor Laboratory, Cold Spring Harbor, New York, latest edition). The reporter gene can be any of a number of standard reporter genes including but not limited to the lacZ gene of *E. coli*, which codes for β -galactosidase. Total genomic DNA is isolated from cells of the pathogen, cleaved with restriction endonucleases to yield fragments of a few hundred base pairs on average. These fragments are then ligated into a unique 30 restriction endonuclease cleavage site at the 5' end of the reporter gene coding region, creating a library of plasmids. The library is then transformed into the pathogen by standard techniques and the resulting transformants are screened for expression of the reporter gene. In the case of lacZ, the transformants can be plated onto medium containing the chromogenic galactosidase substrate X-Gal (5-bromo-4-chloro-3-indolyl-D-galactoside). 35 Transformants that contain a plasmid with an insert carrying a promoter will express β -galactosidase and will turn blue on X-Gal plates. The intensity of the blue color is relative

to the level of expression; promoters of different strength can be selected based on the intensity of the blue color.

The above-described screening procedure can be modified to identify regulated promoters. For example, promoters that are regulated by carbon source availability can be 5 screened on plates that contain different carbon sources. Other modifications are possible and will depend, in part, on the organism in question. To test for species-specificity, the identified promoters are transferred to promoterless reporter plasmids capable of replication and maintenance in a different organism. Truly species-specific or pathogen-specific promoters will not activate the expression of the reporter gene in any other species.

10 Obvious modifications can be used to identify and test artificial promoters composed of synthetic oligonucleotides inserted into the promoterless reporter plasmid.

In one embodiment, the nucleic acids of the invention comprise a tissue-specific promoter operably linked to a sequence encoding a multi-ribozyme of the invention.

The tissue-specific promoter in the ribozyme-producing construct results in tissue-15 specific expression of the ribozyme in tissue(s) that actively transcribe RNA from the selected promoter. Thus, only the target RNA in tissue that utilize the promoter will be cleaved by the ribozyme. Tissue-specific promoters can be used in the present nucleic acid constructs. Examples of these promoters include the binding sites (sequences) for probasin-promoter, a promoter-specific for prostate epithelium prostate-specific antigen 20 (prostate), keratin k4, k13, k7 (epidermal sabaceus glands), albumin (liver), fatty acid binding protein (ilium), whey acidic protein (breast), lactalbumin, smooth muscle actin (smooth muscle), etc.

The pathogen-specific promoter in the ribozyme-producing construct results in pathogen-specific expression of the ribozyme in pathogens or microbes that actively 25 transcribe RNA from the selected promoter. Thus, only the target RNA in pathogens that utilize the promoter will be cleaved by the ribozyme.

It will also be clear that tissue-specific or target-specific promoters not yet identified can be used to target expression of the present ribozymes to the selected tissue(s). Once a target-specific or tissue-specific promoter is identified its binding sequence can be routinely 30 determined by routine methods such as sequence analysis. The promoter is defined by deletion analysis, mutagenesis, footprinting, gel shifts and transfection analyses (Sambrook et al., *supra*).

5.2. EXPRESSION OF THE MULTI-RIBOZYMES IN PROCARYOTIC AND EUKARYOTIC EXPRESSION SYSTEMS

The present invention encompasses expression systems, both eucaryotic and
5 procaryotic expression vectors, which may be used to express the multi-ribozymes of the
present invention.

The invention encompasses the DNA expression vectors and/or viral vectors that
contain any of the foregoing coding sequences operatively associated with a regulatory
element that directs expression of the coding sequences and genetically engineered host
10 cells that contain any of the foregoing coding sequences operatively associated with a
regulatory element that directs the expression of the coding sequences in the host cell. As
used herein, regulatory elements include but are not limited to, inducible and non-inducible
promoters, enhancers, operators and other elements known to those skilled in the art that
drive and regulate expression.

15 The DNA expression vectors and viral vectors containing the nucleic acids encoding
the multi-ribozymes of the present invention may be produced by recombinant DNA
technology using techniques well known in the art. Thus, methods for preparing the
expression vectors and viral vectors of the invention by expressing nucleic acid containing
multi-ribozyme sequences are described herein. Methods which are well known to those
20 skilled in the art can be used to construct expression vectors containing epitope gene
product coding sequences and appropriate transcriptional and translational control signals.
These methods include, for example, *in vitro* recombinant DNA techniques, synthetic
techniques, and *in vivo* genetic recombination. See, for example, the techniques described
25 in Sambrook et al., *supra*. Alternatively, RNA capable of encoding glycoprotein epitope
gene product sequences may be chemically synthesized using, for example, synthesizers.
See, for example, the techniques described in "Oligonucleotide Synthesis", 1984, Gait, M.J.
ed., IRL Press, Oxford, which is incorporated by reference herein in its entirety. Gene
expression may be regulated by a variety of methods known in the art including but not
30 limited to those presented in Mizuno, T. et al., 1984, Proc. Natl. Acad. Sci USA.
81(7):1966070.

5.2.1 EUKARYOTIC AND PROCARYOTIC EXPRESSION VECTORS

The present invention encompasses expression systems, both eucaryotic and
procaryotic expression vectors, which may be used to express the multi-ribozymes. A
35 variety of host-expression vector systems may be utilized to express the selected multi-
ribozyme of the invention. Such host-expression systems represent vehicles by which the

sequences encoding the multiple ribozymes may be introduced into cells and tissues both *in vivo* and *in vitro* but also represent cells which may, when transformed or transfected with the appropriate nucleotide coding sequences, express the multi-ribozymes of the invention. These include but are not limited to microorganisms such as bacteria (e.g., *E. coli*, *B. subtilis*) transformed with recombinant bacteriophage DNA, plasmid DNA or cosmid DNA expression vectors containing selected multi-ribozyme coding sequences; yeast (e.g., *Saccharomyces*, *Pichia*) transformed with recombinant yeast expression vectors containing the selected multi-ribozyme coding sequences; insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus) containing the selected multi-ribozyme coding sequences; plant cell systems infected with recombinant virus expression vectors (e.g., cauliflower mosaic virus, CaMV; tobacco mosaic virus, TMV) or transformed with recombinant plasmid expression vectors (e.g., Ti plasmid) containing selected multi-ribozyme coding sequences; or mammalian cell systems (e.g., COS, CHO, BHK, 293, 3T3) harboring recombinant expression constructs containing promoters derived from the genome of mammalian cells (e.g., metallothionein promoter) or from mammalian viruses (e.g., the adenovirus late promoter; the vaccinia virus 7.5K promoter).

5.3 DELIVERY AND EXPRESSION SYSTEMS

Until the discoveries of present invention, the therapeutic use of ribozymes in eukaryotes was limited because a convenient and efficient delivery system has not been available. A key to the present invention is the strategies used to deliver the ribozymes to the targeted microorganism. Two separate classes of delivery systems can be manufactured, one biologic in nature and the other abiologic.

The key features of the present invention are the combination of ribozymes with viral delivery and assembly of the virions using a unique combination of plasmid features.

5.3.1 ABIOLOGIC DELIVERY VEHICLES

Abiologic delivery of the ribozymes is accomplished by a variety of methods, including packaging plasmid DNA carrying the nucleic acids that code for the ribozyme(s) 30 into liposomes or by complexing the plasmid DNA carrying the nucleic acids that code for the ribozyme(s) with lipids or liposomes to form DNA-lipid or DNA-liposome complexes. The liposome is be composed of cationic and neutral lipids commonly used to transfet cells *in vitro*. The cationic lipids complex with the plasmid DNA and form liposomes.

A liposome is provided, comprising a nucleic acid encoding a multi-ribozyme of the invention. A liposome is provided, comprising a nucleic acid comprising a tissue-specific promoter or target-RNA specific pathogen-specific promoter upstream from a sequence

encoding multi-ribozyme of the invention. For example, a liposome is provided, comprising a nucleic acid comprising a pathogen-specific promoter upstream from a sequence encoding a triple ribozyme comprising a) a 5' autocatalytically cleaving ribozyme sequence, b) a catalytic ribozyme comprising a target RNA-specific binding site and c) a 3' 5 autocatalytically cleaving ribozyme sequence. A liposome is provided, comprising a nucleic acid comprising a tissue-specific promoter or target-RNA specific pathogen-specific promoter upstream from a sequence encoding a) a 5' autocatalytically cleaving ribozyme sequence and/or a 3' autocatalytically cleaving ribozyme; and b) catalytic ribozyme(s) comprising one or more target RNA-specific trans-acting ribozymes. The liposome of the 10 invention, wherein the nucleic acid encodes more than one triple ribozyme or ribozyme cassette is provided. The liposome can comprise any ribozyme-encoding nucleic acid, particularly those described herein.

The liposomal delivery systems of the invention can be used to deliver a nucleic acid comprising a pathogen-specific promoter upstream from a sequence encoding a triple 15 ribozyme comprising a) a 5' autocatalytically cleaving ribozyme sequence, b) a catalytic ribozyme comprising one or more target RNA-specific binding site and c) a 3' autocatalytically cleaving ribozyme sequence. The liposomal delivery systems of the invention can be used to deliver a nucleic acid comprising a pathogen-specific promoter upstream from a sequence encoding any of the multi-ribozymes described herein. The 20 multi-ribozyme that is administered to a subject can further comprise a liposome.

Cationic and neutral liposomes are contemplated by this invention. Cationic liposomes can be complexed with the a negatively-charged biologically active molecule (e.g., DNA) by mixing these components and allowing them to charge-associate. Cationic liposomes are particularly useful when the biologically active molecule is a nucleic acid 25 because of the nucleic acids negative charge. Examples of cationic liposomes include lipofectin, lipofectamine, lipofectace and DOTAP (Hawley-Nelson et al., 1992, *Focus* 15(3):73-83; Felgner et al., 1992, *Proc. Natl. Acad. Sci. U.S.A.* 84:7413; Stewart et al., 1992, *Human Gene Therapy* 3:267-275). Procedures for forming cationic liposomes encasing substances are standard in the art (Nicolau et al., 1987, *Methods Enzymol.* 30 149:157) and can readily be utilized herein by one of ordinary skill in the art to encase the complex of this invention.

In yet another embodiment of the present invention, the plasmid DNA carrying the gene(s) or nucleic acids that codes for the ribozymes of the invention are complexed with liposomes using an improved method to achieve increased systemic delivery and gene 35 expression (Templeton et al., 1997, *Nature Biotechnology* 15: 647-652). In accordance with the present invention, an improved formulation of cationic lipids which greatly

increase the efficiency of DNA delivery to host cells, with extended half-life *in vivo* and procedures to target specific tissues *in vivo*. For example, but not by limitation, peptides and proteins may be engineered to the outer lipid bilayer, such as liver specific proteins, leads to substantially enhanced delivery to the liver etc.

5 In one embodiment of the present invention, systemic delivery and *in vivo* and *ex vivo* gene expression is optimized using commercially available cationic lipids, *e.g.*, dimethyldioctadecylammonium bromide (DDAB); a biodegradable lipid 1, 2-bis(oleoyloxy)-3-(trimethylammonio) propane (DOTAP); these liposomes may be mixed with a neutral lipid, *e.g.*, L- α dioleoyl phosphatidylethanolamine (DOPE) or cholesterol (Chol), two
10 commonly used neutral lipids for systemic delivery. DNA:liposome ratios may be optimized using the methods used by those of skill in the art (*see e.g.*, Templeton et al., *supra*).

In yet another embodiment of the present invention, the plasmid DNA carrying the genes or nucleic acids encoding the ribozymes of the invention may be delivered via
15 polycations, molecules which carry multiple positive charges and are used to achieve gene transfer *in vivo* and *ex vivo*. Polycations, such as polyethylenimine, may be used to achieve successful gene transfer *in vivo* and *ex vivo* (*see e.g.*, Boletta et al., 1996, *J. Am. Soc. Nephrol.* 7:1728).

The liposomes may be incorporated into a topical ointment for application or
20 delivered in other forms, such as a solution which can be injected into an abscess or delivered systemically.

Plasmid DNA coding for the ribozymes is used rather than preformed ribozymes for the following reasons. Plasmid DNA allows the targeted cells to produce the ribozyme and, thus, results in a higher delivered dose to the cell than can be expected by delivery of
25 ribozyme RNA via liposome. The DNA also provides specificity of action based on target sequence specificity. The liposomes deliver their DNA to any cell in the area of administration, including cells of the host. The promoter driving the transcription of the ribozyme is specific for the targeted microorganism and, thus, will be inactive in other cell types. Therefore, liposomal delivery of DNA coding for the ribozyme provides
30 amplification and specificity.

5.3.2 BIOLOGIC DELIVERY VEHICLES

Not all microorganisms are expected to take up DNA delivered by liposome. Consequently, a biologic delivery system is also required. The biologic delivery vehicle of
35 the multi-ribozyme or ribozymes of the invention takes advantage of the fact that generalized transducing particles completely lack DNA originating from the viral vector.

Instead such particles only contain sequences of host origin. Consequently, the invention uses a biologic assembly of viral head proteins (packaging elements for the antimicrobial therapeutic) around the nucleic acid containing the necessary genetic elements that will insure the desired level of expression of the ribozyme(s).

5

5.3.2.1 EXPRESSION OF MULTI-RIBOZYMES IN RECOMBINANT VIRAL VECTORS

In another embodiment of the present invention, either a live recombinant viral vaccine or an inactivated recombinant viral vector expressing the selected multi-ribozyme can be engineered. In this regard, a variety of viruses may be genetically engineered to express the selected multi-ribozymes. For gene therapy purposes, it may be required that the recombinant viruses display attenuation characteristics. Current live recombinant virus candidates for use in humans are either cold adapted, temperature sensitive, or attenuated. The introduction of appropriate mutations (e.g., deletions) into the templates used for transfection may provide the novel viruses with attenuation characteristics. For example, specific multiple missense mutations that are associated with temperature sensitivity or cold adaptation can be made into deletion mutations and/or multiple mutations can be introduced into individual viral genes. These mutants should be more stable than the cold or temperature sensitive mutants containing single point mutations and reversion frequencies should be extremely low. Alternatively, recombinant viruses with "suicide" characteristics may be constructed. Such viruses go through only one or a few rounds of replication in the host.

In accordance with the present invention, a wide variety of viruses and viral vectors may be used to deliver the nucleotide sequences encoding the multi-ribozymes of the present invention, a few examples of which are described below.

25 Retroviral vectors are commonly used to deliver genes to host cells both *in vivo* and *ex vivo*. Retroviral vectors are extremely efficient gene delivery vehicles that cause no detectable harm as they enter the cells. The retroviral nucleic acid may integrate into host chromosomal DNA allowing for long-term persistence and stable transmission to future progeny, such a vector would be useful for the delivery of a multi-ribozyme used to target a 30 cellular gene product involved in a chronic or hereditary disorder or to target a viral gene or a microbial gene or a parasitic gene involved in a chronic or persistent infection. An example of an appropriate retroviral vector are, lentiviruses which have the advantage of infecting and transducing non-dividing cells. In such an embodiment, a lentiviral vector 35 encoding a packagable RNA vector genome and operably linked to a promoter in which all the functional retroviral auxiliary genes are absent, is used to transfer the DNA encoding the multi-ribozyme of the present invention. Examples of such vectors are described in WO

98/17815, WO 98/17816 and WO 98/17817, each of which is incorporated herein by reference in their entirety.

In yet another embodiment, non-integrating viral vectors which infect and transduce non-dividing cells, such as adenoviral vectors may be used to deliver the multi-ribozymes 5 of the present invention. Adenoviral vectors have several advantages because it avoids risks associated with permanently altering the host cell genome or of promoting insertional mutagenesis. Adenoviruses are one of the best developed non-integrating viral vectors and can be used to transfer expression cassettes of up to 75 kb. Recombinant adenoviruses can be produced at very high titers is highly infectious and efficiently transfer genes to a wide 10 variety of non-replicating and replicating cells and is ideal for *in vivo* mammalian gene transfer.

Adenovirus-based vectors are relatively safe and can be manipulated to encode the desired multi-ribozyme and at the same time to be inactivated in terms of their ability to replicate in a normal lytic viral life cycle. Adenovirus has a natural tropism for airway 15 epithelia. Therefore, adenovirus-based vectors are particularly preferred for respiratory gene therapy applications. In a particular embodiment, the adenovirus-based gene therapy vector comprises an adenovirus 2 serotype genome in which the Ela and the Elb regions of the genome, which are involved in early stages of viral replication have been deleted and replaced by nucleotide sequences of interest. In a further embodiment, the adenovirus - 20 -based gene therapy vector contains only the essential open reading frame (ORF3 or ORF6 of adenoviral early region 4 (E4) and is deleted of all other E4 open reading frames, or may additionally contain deletions in the E3 regions (see e.g. U.S. Patent No. 5,670,488, incorporated herein by reference in its entirety). In another embodiment, the adenovirus-based therapy vector used may be a pseudo-adenovirus (PAV), which contain no harmful 25 viral genes and a theoretical capacity for foreign material of nearly 36 kb.

In another embodiment, adeno-associated virus (AAV) systems may be used to deliver the multiple ribozymes of the present invention. AAV has a wide host range and AAV vectors have currently have been designed which do not require helper virus.

Examples of such AAV vectors are described in WO 97/17458.

30 Vaccinia viral vectors may be used in accordance with the present invention, as large fragments of DNA are easily cloned into its genome and recombinant attenuated vaccinia variants have been described (Meyer, et al., 1991, J. Gen. Virol. 72:1031-1038). Orthomyxoviruses, including influenza; Paramyxoviruses, including respiratory syncytial virus and Sendai virus; and Rhabdoviruses may be engineered to express mutations which 35 result in attenuated phenotypes (see U.S. Patent Serial No. 5,578,473, issued November 26, 1996 incorporated herein by reference in its entirety). These viral genomes may also be

engineered to express foreign nucleotide sequences, such as the selected multi-ribozymes of the present invention (see U.S. Patent Serial No. 5,166,057, issued November 24, 1992 incorporated herein by reference in its entirety). Reverse genetic techniques can be applied to manipulate negative and positive strand RNA viral genomes to introduce mutations 5 which result in attenuated phenotypes, as demonstrated in influenza virus, Herpes Simplex virus, cytomegalovirus and Epstein-Barr virus, Sindbis virus and poliovirus (see Palese et al., 1996, Proc. Natl. Acad. Sci. USA 93:11354-11358). These techniques may also be utilized to introduce foreign DNA, i.e., the selected multi-ribozymes, to create recombinant 10 viral vectors to be used in accordance with the present invention. In addition, attenuated adenoviruses and retroviruses may be engineered to express the multi-ribozymes.

Therefore, a wide variety of viruses may be engineered to design the multi-ribozyme delivery vehicles of the present invention.

The viral vectors of the present invention may be engineered to express the multi-ribozymes in a tissue-specific manner. For example, the promoter of the carcinoembryonic 15 antigen (LEA) is expressed in a proportion of breast, lung and colorectal cancers, but rarely in healthy tissues. In order to target a hepatoma, the α -fetoprotein (AFP) promoter whose activity is restricted to malignant cells. Proliferating cells can be targeted with a flt-1 promoter, which has been shown to allow preferential targeting of proliferating endothelial 20 cells. See Miller et al., 1997, Human Gene Therapy 8:803-815.

The virion of the present invention can also be any bacteriophage which specifically 25 infects a bacterial pathogen of the present invention as well as any virus which can be specifically targeted to infect the pathogen of the present invention (Soothill, J.S., 1992, J. Med. Microbiol. 37:358-261).

For example, the bacteriophage can include, but is not limited to, those specific for 30 bacterial cells of the following genera: *Bacillus*, *Campylobacter*, *Corynebacterium*, *Enterobacter*, *Enterococcus*, *Escherichia*, *Klebsiella*, *Mycobacterium*, *Pseudomonas*, *Salmonella*, *Shigella*, *Staphylococcus*, *Streptococcus*, *Vibrio*, *Streptomyces*, *Yersinia* and the like (see e.g., the American Type Culture Collection Catalogue of Bacteria and Bacteriophages, latest edition, Rockville, MD), as well as any other bacteriophages now known or later identified to specifically infect a bacterial pathogen of this invention.

In the virions of the present invention, the non-viral DNA can encode any ribozyme of the invention. In the virions of the non-viral DNA can comprises a pathogen-specific promoter upstream from a sequence encoding one or more ribozyme cassettes containing one or more trans-acting ribozymes with in the catalytic core(s). In one example the 35 multi-ribozyme may comprise a) a 5' autocatalytically cleaving ribozyme sequence, b) a

catalytic ribozyme comprising two target RNA-specific binding site sequences and c) a 3' autocatalytically cleaving ribozyme sequence.

The virions of the present invention within the non-phage DNA which encodes more than one triple ribozyme is also provided. There are several options for constructing the 5 multi-ribozyme encoding sequences: 1) ribozymes directed to different targets in the same pathogen; 2) multiple copies of the same ribozyme; and 3) multiple ribozymes directed to multiple targets. These may be combined in various ways, *e.g.*, multiple copies of DNA encoding 4 different ribozymes in a single construct under one promoter. The promoter can have the chosen level of specificity as described herein.

10 The virion can contain a nucleic acid encoding at least two different ribozyme cassettes. The virion can contain a nucleic acid encoding more than one copy of a ribozyme cassette. The virion can comprise any ribozyme-encoding nucleic acid, particularly those described herein.

15 The nucleic acid delivered by the virion or liposome can encode more than one ribozyme cassette. The nucleic acid can encode at least one or more different ribozyme cassettes. The nucleic acid can encode more than one copy of the same ribozyme cassette. The nucleic acid can encode a cassette in which the cassette is a triple ribozyme. Each 20 ribozyme cassette can contain one or more trans-acting ribozymes. Trans-acting ribozymes can be directed to the same or different targets. The nucleic acid can encode combinations of different ribozymes, some or all of which may be encoded in more than one copy.

Additionally, the ribozymes may be linked by a short spacer. The ribozymes of the invention can be stabilized by a hairpin loop.

This delivery system consists of a DNA plasmid carrying the gene(s) coding for the 25 ribozyme(s) packaged into viral particles. Specificity is conferred by the promoter driving transcription of the ribozymes and by the host specificity of the viral vehicle. The invention provides examples of the system using bacteriophage lambda virions to package DNA carrying ribozymes directed against *Escherichia coli*. Similar strategies are used to generate Multi-ribozyme capable of delivering ribozymes directed against other 30 microorganisms. The virions used to package the DNA can be species specific, such as the virion derived from the bacteriophage lambda coat, or they can possess a broader host range, such as virion derived from bacteriophage P1. Broad host-range viruses facilitate production of the anti-microbial agents without the loss of species specificity because species-specific promoters are used to direct the transcription of the ribozymes which are 35 directed against species specific targeted RNA sequences.

One example of construction the present multi-ribozyme invention entails the use of a plasmid carrying the ribozyme gene(s), a plasmid origin of replication, a selectable marker for plasmid maintenance, the minimal lambda origin of replication, and cos sites, which are required for packaging of DNA into lambda virions. This plasmid is maintained in a lambda lysogen that is defective in integration/excision and recombination functions. The defective lysogen provides all of the replication factors needed to activate the lambda origin of replication on the plasmid and all of the structural components needed to form mature virions; however, the lysogen is not able to replicate and package its own DNA into the virions. The lysogen also carries the cI857 temperature-sensitive repressor mutation.

5 Induction of the lysogen is described in the Examples. A similar strategy can be used to generate ribozyme-encoding plasmids packaged into bacteriophage P1 virions.

10 A common bacteriophage of *E. coli*, P1, is an attractive delivery vehicle for a multi-ribozyme for a number of reasons. First and foremost, P1 has a broad intergenera and interspecies range (Yarmolinsky, M.B., and N. Steinberg, 1988, *Bacteriophage, P1*, p.291-15 438. In R. Calendar (ed.), *The Bacteriophages*, vol. 1, Plenum Press. New York). The P1 receptor of *E. coli* is the terminal glucose of the lipopolysaccharide (LPS) core lysogenic ring of the bacterial outer membrane (Masters, M., 1966, *Generalized Transduction*, p. 2421-2441. In F. Neidhardt (ed.), *Escherichia coli and Salmonella: Cellular and Molecular Biology*, 2d ed. Vol.2, ASM Press, Washington, D.C.). Yarmolinsky and Sternberg report 20 that in addition to *E. coli*, this particular phage has the ability to inject its nucleic acid into a large number (>25) of diverse gram negative bacteria (Yarmolinsky, M.B., and N. Steinberg, *supra*) Secondly, P1 can accommodate a significant amount of genetic information, over 2% (100,000 bp) of the DNA of *E. coli* (Masters, M., *supra*). Consequently, gene dosage of the ribozymes can be increased through multiplication of the 25 present ribozyme cassettes, thereby increasing the microbicidal activity of the multi-ribozyme. Bacterial strains already exist that can be readily modified to package ribozyme coding DNA *in vivo* by a process similar to that described above. Additionally, a process utilizing *in vitro* packaging is also possible. *in vitro* packaging can be accomplished through the addition of PAC-sites to the genetic information already present within the ribozyme 30 construct. P1 packaging initiates within one of the P1 PAC genes (Sternberg, N., and J. Coulby, 1987, *J. Mol. Biol.* 194(3):469-79). It has been reported that the active PAC site is contained within a 161 base-pair segment of the P1 EcoR1 fragment 20 (Sternberg, N., and J. Coulby, *supra*). Thus, the phage head serves as a molecular syringe that delivers the inactivating ribozyme(s) to the pathogen.

5.4 HOST CELLS

The present invention encompasses the expression of the multi-ribozymes in primary cells, animal and insect cell lines for *in vitro* screening assay and *ex vivo* gene therapy. In accordance with the present invention, a variety of primary or secondary cells or 5 cell strains may be used including but not limited to cells isolated from skin, bone marrow, liver, pancreas, kidney, adrenal and neurological tissue to name a few. Other cells types that may be used in accordance with the present invention are immune cells (such as T-cells, B-cells, natural killer cells, etc.), macrophages/monocytes, adipocytes, pericytes, fibroblasts, neuronal cells, reticular cells etc. In a further embodiment, secondary cell lines 10 may be used as engineered responsive cells and tissues in accordance with the present invention, including, but not limited to hepatic cell lines, such as CWSV, NR, Chang liver cells, or other cell lines such as CHO, VERO, BHK, Hela, COS, MDCK, 293, 373, CaSki and W138 cell lines.

For long term, high-yield production of recombinant proteins, stable expression is 15 preferred. For example, cell lines which stably express the selected target epitope may be engineered. Rather than using expression vectors which contain viral origins of replication, host cells can be transformed with DNA controlled by appropriate expression control elements (e.g., promoter sequences, enhancer, sequences, transcription terminators, polyadenylation sites, etc.), and a selectable marker. Following the introduction of the 20 foreign DNA, engineered cells may be allowed to grow for 1-2 days in an enriched media, and then are switched to a selective media. The selectable marker in the recombinant plasmid confers resistance to the selection foci (e.g., by stably integrating the plasmid into their chromosomes) and allows cells to and grow to form which in turn can be cloned and expanded into cell lines. This method may advantageously be used to engineer cell lines. 25 This method may advantageously be used to engineer cell lines which express the selected gene products. Such cell lines would be particularly useful in screening and evaluation of compounds that affect the endogenous activity of the selected gene product.

A number of selection systems may be used, including but not limited to the herpes simplex virus thymidine kinase (Wigler, et al., 1977, Cell 11:223), hypoxanthine-guanine 30 phosphoribosyltransferase (Szybalska & Szybalski, 1962, Proc. Natl. Acad. Sci. USA 48:2026), and adenine phosphoribosyltransferase (Lowy, et al., 1980, Cell 22:817) genes can be employed in tk⁻, hgprt⁻ or aprt⁻ cells, respectively. Also, antimetabolite resistance can be used as the basis of selection for the following genes: DHFR, which confers 35 resistance to methotrexate (Wigler, et al., 1980, Natl. Acad. Sci. USA 77:3567; O'Hare, et al., 1981, Proc. Natl. Acad. Sci. USA 78:1527); gpt, which confers resistance to mycophenolic acid (Mulligan & Berg, 1981, Proc. Natl. Acad. Sci. USA 78:2072); neo,

which confers resistance to the aminoglycoside G-418 (Colberre-Garapin, et al., 1981, *J. Mol. Biol.* 150:1); and hygro, which confers resistance to hygromycin (Santerre, et al., 1984, *Gene* 30:147).

5 **5.5 TARGET SELECTION & TARGET RNA CLEAVAGE**

The complexity of human RNA is about 100 fold lower than that for human DNA, and specificity can be achieved with as few as 12-15 base pairs. The stability of the RNA-RNA duplex is effected by several factors, such as GC content, temperature, pH, ionic concentration, and structure. The nearest neighbor rules can provide a useful estimate of the 10 stability of the duplex (Castanotto et al., 1994, *Advances in Pharmacol.* 25:289-317).

The catalytic ribozyme of the invention also includes a catalytic sequence, which cleaves the target RNA near the middle of the site to which the target RNA-specific binding site (sequences) bind. In the hammerhead-type of ribozyme, the catalytic sequence is generally highly conserved (Bertrand E., et al., 1994, *Nucleic Acids Research* 22(3) :293-15 300; Inokuchi, Y., N. et al., 1994, *J. biol. Chem.* 269(15):11361-6). The conserved catalytic core residues are 5' CUGANGA 3' and 5' GAAA 3' linked by an evolutionarily conserved stem-loop structure.

The most conserved and probably most efficiently cleaved sequence on the target RNA is 5' GUC 3'. However, NUX (wherein X = A, U or C) can also be cleaved 20 efficiently. Such cleavage sites are ubiquitous in most RNAs allowing essentially all RNA's to be targeted (Whitton, J. Lindsay "Antisense Treatment of Viral Infection" *Adv. in Virus Res.* Vol. 44, 1994).

With regard to the selection of the appropriate sites on target RNA, it is known that target site secondary structure can have an effect on cleavage *in vitro* (Whitton, 1994, 25 *supra*). A number of procedures are available to select accessible sites in RNA targets. In a preferred procedure, a library screen may be employed to select appropriate sites on the target RNA. Accessibility of the selected site may then be confirmed using techniques known to those skilled in the art. Thus, the selected target molecule's sequence can be routinely screened for potential secondary structure, using the program RNAFOLD (from 30 the PCGENE group of programs or available on the Internet). Thus, reasonable predictions of target accessibility can be made. Computer assisted RNA folding (Castanotto et al., 1994, *supra*), along with computational analysis for 3-dimensional modeling of RNA (Major et al., *Science* 253:1255-1260, 1991 and Castanotto et al., 1994, *supra*) is certainly effective in guiding the choice of cleavage sites.

35 The multi-ribozymes of the present invention may be engineered to target a wide variety of cellular RNAs, tumor or cancer associated with RNAs, parasitic RNA etc. The

internal ribozyme can be targeted to noncellular RNAs necessary for growth of parasites, virus life cycles, etc., and expression can be driven with tissue-specific or virus-specific promoters.

The first critical component in the assembly of the multi-ribozyme is the selection of appropriate RNA targets. For ribozymes to be effective anti-microbial therapy, it is preferable to target the RNA of, for example, several key or essential proteins, tRNA, rRNA or any other RNA molecule essential for cell viability or fitness, in order to insure complete inactivation and prevent escape of the invading microorganism. For example, four bacterial genes, essential for viability and unrelated in activity, have been selected and are described herein to highlight how the selection of appropriate mRNA targets is carried out for the preferred construction of the multi-ribozyme against prokaryotic targets. Cross-genera RNA targets can be used to design a multi-ribozyme that can have broad application, modified by the specificity of the promoter.

In one embodiment of the invention, the first ribozyme targets an essential transcription factor, the second ribozyme targets an essential general secretory component, the third ribozyme targets an essential component of the primosome required for DNA biosynthesis and the fourth ribozyme targets an enzyme required for cell division. Consequently, the ribozymes are redundant in the fact that they inhibit growth by specifically targeting a fundamental process required for bacterial growth. Thus, this can minimize the development of resistance to the antimicrobial therapeutic.

Examples of ribozymes targets of the invention include but are not limited to the following:

The first gene, *rpoA*, produces an essential protein, *rpoA* or the alpha subunit of RNA core polymerase. *rpoA* was selected rather than the other components of the RNA polymerase holoenzyme, because it is thought to facilitate the assembly of an active RNA Polymerase enzyme complex. Inactivation of the *rpoA* transcript results in a decrease in the intracellular concentration of the holoenzyme RNA polymerase rendering the cell less able to respond to changes demanded of it once it has invaded a new host. The nucleotide sequence of *rpoA* is known for a large number of microorganisms (>20 genera) and they are readily available from GenBank.

The second ribozyme target can be the mRNA of the *secA* gene from bacteria. The product of this gene is the essential and rate-limiting component of the general secretory pathway in bacteria (Bassford, P., et al., 1991, Cell 65(30):367-368). *secA* has been found in every prokaryotic cell investigated to date. Additionally, its biosynthesis is translationally coupled to the upstream gene, X (Schmidt, M.G., et al., 1991, J. Bacteriol. 173(20):6605-11), presenting a convenient target for a ribozyme. Inhibition or decreased synthesis of *secA*

is also sufficient to confer a reduction in viability to the cell (Schmidt, M.D., and D.B. Oliver, 1989, *J. Bacteriol.* 171(2):643-9). Furthermore, as a pathogen responds to changes required of the infectious process a change in the availability of a key protein such as *secA* will disadvantage the pathogen enabling the host to counteract it. Finally, control over the 5 secretion-responsive expression of *secA* is at the level of translation (Christoffersen, R.E., and J. J. Mann. 1995, *J. Med. Chem.* 38(12):2023-37), and the regulatory sequences within its polycistronic message have been localized to a region comprised of the end of the upstream gene, X, and the beginning of *secA*. Consequently, inactivation of the transcript by the catalytic cleavage of a ribozyme has profound consequences for the viability of the 10 invading microorganism.

The third ribozyme can target an essential factor for DNA biosynthesis, DnaG. Every 1 to 2 seconds, at least 1,000 times for each replication fork within *E. coli*, priming of an Okazaki fragment is repeated as a result of an interaction between the cellular primase DnaG (Bouche, J.P., et al., 1975, *J. Biol. Chem.* 250:5995-6001) and *DnaB* (Marians, K.J. 15 1996, *Replication Fork Propagation*, p. 749-763. In F.C. Neidhardt (ed.), *Escherichia coli and Salmonella: Cellular and Molecular Biology*, 2nd ed, vol. 1. American Society for Microbiology, Washington, D.C.). As would be expected of protein required every 1 to 2 seconds during replication, a lesion within dnaG or an alteration in its concentration results in an immediate stop phenotype (Marians, K.J. 1996, *supra*; Weschler, J.A. and J.D. Gross, 20 1971, *Mol. Gen. Genet.* 113:273-284). Therefore, inactivation of the dnaG message by a ribozyme should have profound cellular consequences in that general priming of the lagging strand is reduced if not eliminated. DnaG is a component of the primosome, a multi-protein complex responsible for priming replication. Any of the components of the primosome, either individually or in any combination, can serve as a target for inactivation of the 25 primosome and, thus, kill the cell. The other components of the primosome are *DnaB*, *DnaC*, *DnaT*, *PriA*, *PriB*, and *PriC*. Thus, the primosome is also sufficiently complex to provide numerous other targets (*DnaB*, *DnaC*, *DnaT*, *PriA*, *PriB* and *PriC*) for inactivation by the trans ribozyme.

The fourth target can be *ftsZ*. This gene also encodes an essential protein, *ftsZ*, that 30 is required for cell division in that it is responsible for the initiation of separation (see e.g., Tetart, F., and J.P. Bouche, 1992, *Mo. Microbiol.* 6(5):615-20; Haseloff, J., and W.L. Gerlach, 1988, *Nature* 334(6183):585-91; Sullivan, S. M., 1994, *J. Invest Dermatol.* 103(5 Supl):858-895). *ftsZ* was selected because its synthesis was under the control of an antisense RNA molecule encoded by the gene *dicE*. Transcription of *dicE* is all that is 35 needed to inhibit the translation of *ftsZ*; thus, overexpression of this antisense molecule is sufficient to cause an inhibition of cell division and a reduction in viability. There is an

advantage of using a ribozyme against *ftsZ* over the antisense molecule, dicE. Specifically, the ribozyme functions catalytically while dicE functions stoichiometrically. Thus, upon cleavage of the *ftsZ* message the ribozyme attacks additional copies of *ftsZ* inhibiting the division of the cell. The nucleotide sequence of *ftsZ* like the other targets selected, is

5 commonly available from GenBank.

Important examples which are specifically presented in the application are:

- A) Use of the albumin promoter with a Hepatitis B virus target (chosen to cleave the viral RNA pregenome, S protein, polymerase/reverse transcriptase, and polymerase/and x protein transcripts using the same ribozyme target site);
- 10 B) Use of generic promoters active in erythrocytes, using a ribozyme targeted to highly conserved regions of the EMP-1 protein family from *P. falciparum*, which are necessary for cytoadherence and antigenic variation in malaria; and
- 15 C) Use of the keratin 7 promoter, with trans-acting ribozymes targeted to a specific sites near the translational start site of the E6 protein, a site known to be critical for expression of both the E6 and E7 proteins which are intimately involved in cervical carcinogenesis, as well as a more 3' site in a highly conserved region of the E6 protein.
- 20 D) Use of the HBV or HPV early promoter, with trans-acting ribozymes targeted to a specific sites near the translational start site of the E6 protein, a site known to be critical for expression of both the E6 and E7 proteins which are intimately involved in cervical carcinogenesis, as well as a more 3' site in a highly conserved region of the E6 protein.
- 25

Examples of bacterial pathogens that can be targeted by the multi-ribozyme construct of the present invention include, but are not limited to, species of the following genera: *Salmonella*, *Shigella*, *Chlamydia*, *Helicobacter*, *Yersinia*, *Bordatella*,
30 *Pseudomonas*, *Neisseria*, *Vibrio*, *Haemophilus*, *Mycoplasma*, *Streptomyces*, *Treponema*,
Coxiella, *Ehrlichia*, *Brucella*, *Pasteurella*, *Clostridium*, *Corynebacterium*, *Listeria*,
Bacillus, *Erysipelothrix*, *Rhodococcus*, *Escherichia*, *Klebsiella*, *Enterobacter*, *Serratia*,
Staphylococcus, *Streptococcus*, *Legionella*, *Mycobacterium*, *Proteus*, *Campylobacter*,
Enterococcus, *Acinetobacter*, *Morganella*, *Moraxella*, *Citrobacter*, *Rickettsia*, *Rochlimeae*
35 and any other bacterial species or genera now known or later identified to be pathogenic.

The pathogen of the present invention can also include, but is not limited to pathogenic species of yeast/fungal genera (e.g., *Candida*, *Cryptococcus*, *Aspergillus*, *Trichophyton*, *Microsporum*) as well as any other yeast or fungus now known or later identified to be pathogenic. Furthermore, the pathogen of the present invention can be a 5 parasite, including, but not limited to, members of the Apicomplexa phylum such as, for example, *Babesia*, *Toxoplasma*, *Plasmodium*, *Eimeria*, *Isospora*, *Atoxoplasma*, *Cystoisospora*, *Hammondia*, *Besniotia*, *Sarcocystis*, *Frenkelia*, *Haemoproteus*, *Leucocytozoon*, *Theileria*, *Perkinsus* and *Gregarina* spp.; *Pneumocystis carinii*; members 10 of the Microspora phylum such as, for example, *Nosema*, *Enterocytozoon*, *Encephalitozoon*, *Septata*, *Mrazekia*, *Amblyospora*, *Ameson*, *Glugea*, *Pleistophora* and *Microsporidium* spp.; and members of the Ascetospora phylum such as, for example, *Haplosporidium* spp., as 15 well as any other parasite now known or later identified to be pathogenic.

Examples of viral pathogens include, but are not limited to, retroviruses (human immunodeficiency viruses), herpes viruses (herpes simplex virus; Epstein Barr virus; 15 varicella zoster virus), orthomyxoviruses (influenza), paramyxoviruses (measles virus; mumps virus; respiratory syncytial virus), picorna viruses (Coxsackie viruses; rhinoviruses), hepatitis viruses (hepatitis C), bunyaviruses (hantavirus; Rift Valley fever virus), arenaviruses (Lassa fever virus), flaviviruses (dengue fever virus; yellow fever virus; chikungunya virus), adenoviruses, birnaviruses, phleboviruses, caliciviruses, 20 hepadnaviruses, orbiviruses, papovaviruses, poxviruses, reoviruses, rotaviruses, rhabdoviruses, parvoviruses, alphaviruses, pestiviruses, rubiviruses, filoviruses, coronaviruses and any other virus now known or later identified to be pathogenic.

The virion construct used in this method can comprise any ribozyme-encoding nucleic acid, particularly those described herein targeted to genes of the pathogen. The 25 virion can be a bacteriophage, or other virus selected for its ability to target a specific cell-type, microorganism or animal. The bacteriophage can be lambda, P1 or other phage. When P1 is the virion, the non-viral DNA can further comprise a PAC site is also provided. This construct is preferred when using P1. Alternatively, the virion can be selected because it has a broad range of targets.

30 The above targeting method, wherein the virion is a bacteriophage is provided. The bacteriophage can be lambda, P1 or other phage. The targeting method, wherein the non-viral DNA further comprises a PAC site is also provided. This construct is preferred when using P1.

It should be clear that any other essential protein of a pathogen can have its message 35 targeted in the present invention, and that determining which proteins are essential can be routinely determined according to standard protocols in the art. In fact, there are over

52,000 viral, 41,000 bacterial and 12,300 fungal sequences deposited in the public section of the Entrez Database at the National Center for Biotechnology Information. Any of these can be used to design the catalytic trans ribozyme of the multi-ribozyme. Thus, multi-ribozyme can comprise ribozymes targeted to these other messages.

5 In addition to targeting mRNA of essential proteins ribozymes may be targeted against other RNA species within the cell. Specifically, appropriate targets in bacteria, fungi and other lower eukaryotes include ribosomal RNA such as Small Subunit RNAs (SSU) or Large Subunit (LSU) and tRNA molecules required for protein synthesis. For example, with respect to pathogenic Staphlococcus, the RNA III moiety is a relatively low
10 abundance transcript which is not translated and should be accessible for cleavage. As long as the RNA targeted contains a canonical ribozyme cleavage domain the multi-ribozyme therapeutic can hybridize and cleave the complementary RNA, thus impacting the fitness of the microbial cell. Additionally, over 3000 rRNA species have been sequenced and aligned. This information is available from the Ribosomal Database Project and should facilitate
15 rapid design and adaptation of ribozyme(s) against such targets. For example the 16S rRNA molecule of bacteria is especially attractive in that there are over 4000 copies of the 16S rRNA per cell. Consequently, a reduction in number slows the process of protein synthesis in so far as the 16S rRNA molecule is involved in the process of translational initiation. Thus, a multi-ribozyme containing ribozymes directed against mRNA and rRNA
20 impacts the fitness of the offending microorganism.

5.6 SCREENING FOR NOVEL TRANS-ACTING RIBOZYMES AND TARGETS

The present invention encompasses the expression of the multi-ribozymes in
25 primary cells, animal and insect cell lines for *in vitro* screening assay and *ex vivo* gene therapy. The present invention also relates to library screening that allows the identification of potential trans-acting ribozymes that may be used in the present invention. For example, the present invention provides *in vivo* hammerhead ribozyme library screen, as outlined in Figure 9. Generally, the hammerhead ribozyme library screen delivers a ribozyme library to
30 host cells (such as prokaryotic/bacterial cells) which expresses the library upon induction (e.g., by methods known in the art). Any clones which carry a lethal ribozyme construct are detected by differences in, or lack of, the clone on a replica plate. The clone is then recovered, the ribozyme is characterized, and the RNA target identified.

This procedure is accomplished by a combination of several techniques known in
35 the art. The plasmid vector which is constructed to contain a regulated promoter. In a preferred embodiment, the regulated promoter will have tight repression in the host cell in

the uninduced state which allows for maintenance of the potentially lethal plasmid within the host. Upon induction (or de-repression of the promoter) a large excess of transcribed product is expressed. When the expressed product (e.g., trans-acting ribozyme) is a sequence for a highly toxic, host-specific ribozyme, the host cell will die.

5 The library itself is a hammerhead ribozyme structure with degenerate bases for the two antisense arms (helix I and III). In a preferred embodiment, the library contains all ribozymes capable of binds to and cleaving any stretch of accessible RNA sequence containing the NUX motif. The library is constructed from a custom synthesized single stranded oligonucleotide. A second strand is synthesized in vitro utilizing a compatible 10 primer and polymerase. The double-stranded oligonucleotide is trimmed with restriction endonucleases to allow for directional cloning into the plasmid vector (such as the modified pChop cassette vector) prepared with compatible cohesive ends. In one embodiment, the plasmid vector utilizes the modified pChop expression cassette which releases the trimmed, transacting, internal ribozyme in an autocatalytic manner upon transcription. The modified 15 pChop also provides a 3' hairpin loop on the internal ribozyme to protect it from exonuclease attack. Additionally, a lacI gene may be cloned onto the plasmid vector to allow for tighter repression of lac operator containing modified pChop expression promoters. In other embodiments of the invention the screening cassette is pClip, pChop, or pSnip cassette. In still other embodiments of the invention, more than one cassette is used.

20 One technique for detecting the lethal events utilizes replica plating. The library is ligated into the appropriate plasmid vector and this ligation is transformed into the bacterial host cells. In one embodiment of the invention, the host cells have a ribosome deficiency such that they are slow-ribosome host cells. The cells are plated onto solid media with the appropriate antibiotic selection and allowed to grow until small, isolated colonies are 25 present. The dish is replica plated using sterile velvet onto solid media containing the inducer compound. Ribozyme expression is then induced in the replica plate colonies (one ribozyme sequence per clonal colony). Replica plates are incubated for approximately equivalent time as the parent plate was originally allowed to incubate, so that colony sizes of the parent plate and replica plate are similar. When the ribozyme is lethal or toxic to the 30 cell, the toxic or lethal ribozyme-containing colony will not grow on the replica plate. Toxic or lethal ribozyme-containing colonies are detected by comparison of the two plates (parent and replica). The original colony corresponding to a lethal ribozyme can be picked and propagated from the original parent plate by standard techniques in the art. The sequence of the ribozyme of the colony is then determined by standard methods in the art. 35 The RNA target may also be determined by complementation. Once sequenced, ribozymes

revealed in the screen may be redesigned and refined to enhance the activity of the ribozymes.

The ribozyme screen provides two important advantages. First, it demonstrates whether particular hammerhead ribozymes can be used as a lethal agent in the given host species. The ribozyme library represents approximately 6.7×10^7 unique hammerhead ribozyme sequences. Following screening of a significant number of colonies (and thus ribozyme sequences), should no lethal events be detected, then the single trans-acting ribozyme approach may be less preferred for the given species. Importantly, however, the expression and screening system is preferably optimized and validated for the host species under investigation. For example, as described in the Section 6, herein, in an *E. coli* host, the UPRIP (LEASHI) promoter expressing the modified pChop cassette coupled with AP LacI element on a pBluescript plasmid backbone gives the highest ratio of induced versus repressed expression levels when assayed in bacterial S03831slow-ribosome cells.

Another important advantage of the screening system is when a positive colony is detected, the ribozyme is known to be lethal in an *in vivo* system. Thus there is decreased discrepancy due to differences arising from *in vitro* to *in vivo* systems. Thus, the *in vivo* screening methods directly demonstrate a susceptible target as well as an effective lethal trans-acting ribozyme.

20 5.7 MULTI-RIBOZYME DESIGN

The multi-ribozyme ribozyme possesses sufficient catalytic activity to inactivate the RNA of the targeted RNAs. From an antimicrobial perspective, hammerhead-type ribozymes are especially attractive since the molecule inactivates gene expression catalytically through the cleavage of the phosphodiester bond of the mRNA. Furthermore, hammerhead-type ribozymes have been re-engineered to function in an intermolecular or transducer (trans) acting state (Haseloff et al., 1988, *Nature* 334(6183):585-91; Uhlenbeck. O.C., 1987, *Nature* 328(6131):59). The catalytic activity of the ribozyme requires a sufficient concentration of the divalent cation, Mg^{+2} , and substrate. The substrate can have any sequence as long as the cleavages site contains the recognition element NUX, where N represents any nucleotide, U corresponds to uracil, and X is any nucleotide except G (Koizumi et al., 1989, *Nucleic Acids Research*. 17(17):7059-71). Ribozymes have been widely demonstrated to function *in vivo* (Christoffersen et al., 1995, *J. Med. Chem.* 38(12):2023-37; Inokuchi et al., 1994, *J. Biol. Chem.* 269(15):11361-6). The present invention improves the initial design of hammerhead-type ribozymes (Taira et al., 1991, *NAR* 19(9):5125-5130) by constructing multi-ribozymes consisting of ribozyme cassettes. Ribozyme cassettes contain one or more *cis*-acting hammerhead ribozymes flanking a

ribozyme that inactivates the targeted RNA(s) as well as one or more flanking sequences. Upon transcription the targeted ribozyme is released as a 60-70 base transcript which not only improves its specificity by reducing non-specific interactions but also improves its catalytic activity as well. This invention includes modifications to and use of the ribozyme 5 described in U.S. Serial No. 08/554,369 and PCT publication No. WO98/24925, which are incorporated by reference herein in their entirety.

The ribozymes of the present invention have several important modifications. The arms of the cis-acting ribozymes have been lengthened by 20 bases. The sequence has been modified to enhance the catalytic activity of the cis-acting elements, for example, those 10 shown in SEQ ID NOS:18-38. Additional restriction sites are included that facilitate easier cloning and manufacturing. Specifically, restriction enzymes have been chosen which produce cohesive ends, so that internal trans-acting ribozymes can be cloned into the ribozyme cassettes in a single vector facilitating construction of targeted ribozymes. Design on the internal ribozymes may also include the use of a hairpin loop at the 3'end of the 15 internal ribozymes. In several embodiments, tRNA elements are present in the 3' end of the multi-ribozyme. The addition of the tRNA elements creates additional structure that improves the stability of multi-ribozyme helping it resist nuclease attack. An inverted nucleotide repeat has been inserted into the 3' end of the multi-ribozyme. The addition of the inverted repeat, a hairpin loop structure, improves the stability of multi-ribozyme, 20 helping it resist nuclease attack (see e.g., Pace, N.R., and D. Smith, 1990, J. Biol. Chem. 256(7):3587-90; Schmidt, M., and N. Delihas, 1995, .FEMS Microbiol. Lett. 133(3) :209-13).

5.8 PROTECTION OF RIBOZYME-PRODUCING CELLS

25 The genes or nucleic acids coding for the ribozymes can be toxic to the cells that are needed to produce the ribozyme-carrying virions. When using a broad host-range virus like P1, the organism used to produce the multi-ribozyme can be different from the target organism. In this way, the producing strain is resistant to the toxic effects of the ribozymes because the ribozymes are not efficiently expressed in the producing strain, due to species- 30 specific promoter elements, and the ribozymes will not have any target RNA molecules to attack, due to the species-specific sequences that target the ribozymes. When using a species-specific virus that must be expressed and assembled within a strain of the targeted microorganism, this toxicity becomes a significant concern. The assembly of a multi-ribozyme consisting of anti-*E. coli* ribozyme genes packaged in lambda will illustrate the 35 approach used to circumvent the toxicity. The ribozymes directed against RNA species of *E. coli* is expressed from a artificial promoter containing consensus promoter elements. This

promoter provides high level transcription of the ribozyme immediately upon infection of targeted cells. In order to prevent the unwanted death of the producing strain of *E. coli*, transcription is repressed in the producing strain by a mechanism not available to the wildtype strains that are targeted for killing. Sequences constituting the DNA binding sites 5 for a heterologous transcription factor are interspersed between the essential activating elements of the ribozyme promoter. Expression of the heterologous transcription factor in the producing strain results in the occlusion of the activating promoter elements and preventing the binding of RNA polymerase. As an example, the gene for the *Saccharomyces cerevisiae* transcription factor Ste12p may be expressed in *E. coli* and bind 10 to its binding sites, the pheromone response element, located within the ribozyme promoter. Ste12p will not be found in wild strains of *E. coli*; therefore, the ribozyme promoter will be accessible to RNA polymerase following delivery of the plasmid to the targeted cells.

An alternative strategy that can protect the producing strain from the toxicity of the ribozymes employs ribozyme-resistant versions of the targeted RNA molecules. This 15 strategy can be used when the target RNA molecule codes for a protein. The ribozyme target site within the mRNA molecule is mutated by site-directed mutagenesis such that the amino acid sequence of the translated protein does not change but the mRNA sequence no longer serves as a substrate for the ribozyme. For example, hammerhead ribozymes require 20 an NUX sequence within the target mRNA for cleavage to occur. By changing this sequence to something else, the ribozyme will not cleave the mRNA. This type of ribozyme resistant version of the target RNA can be expressed from a plasmid or integrated into the chromosome of the producing strain and thus render this strain resistant to the toxic effects 25 of the ribozyme.

Merrill and co-workers reported on the selection of long-circulating bacteriophages 25 as anti-bacterial agents (Merril, C.R. et al., 1966, Proc. Natl. Acad. USA. 93:3188-3192). They were able to show that it is possible to select for phage variants that remain refractory 30 to clearance by the reticuloendothelial system for a period of time sufficient to confer a therapeutic response within an infected animal (Merril, C., *supra*). Specifically, the phage will promptly induce neutralizing antibodies interfering with the phage's ability to attack 35 against the bacteria and opsonins that will restore the vulnerability of the phage (Lederberg, J. ,1966, Smaller fleas . . . ad infinitum: Therapeutic bacteriophage redux. Proc. Natl. Acad. Sci. USA 93:3167-3168.). The improvement in the present invention is that a non-replicative delivery system has an advantage in that once the phage coat has injected the nucleic acid into the targeted bacterium, the expression of the multi-ribozyme ribozyme will 40 destroy the microbe, as opposed to a lytic infection cycle typical of an intact bacteriophage. Consequently, amplification of the phage coat will not be an issue and it is less likely that 45

the non-replicative phage delivery system will generate an immune response such that subsequent use of the delivery system would be jeopardized. Moreover, if the patient has been exposed to a resistant pathogenic microbe and the multi-ribozyme is effective and neutralizes the invading microbe, then it is expected that the microbial antigens liberated as 5 a result of the action of the multi-ribozyme, will illicit sufficient humoral immunity and cell-mediated immunity to confer protection against subsequent attacks.

5.9 ADMINISTRATION

A method of delivering a ribozyme to a target (e.g., a pathogen) in a subject is 10 provided, comprising a) generating a liposome comprising a promoter and ribozyme-encoding sequence; and b) delivering the liposome to the subject, whereby the target-specific promoter directs transcription of the ribozyme in the cells of the target. The target can be a pathogen, for example, a bacteria, fungus, yeast, parasite, virus or non-viral pathogen.

15 A method of targeted delivery of a ribozyme to a pathogen in a subject, comprising a) generating a virion comprising non-viral DNA of the invention; b) combining it with a liposome; and b) delivering the liposome containing the virion to the subject, whereby liposome enters the eukaryotic cell and releases the virion, which delivers the DNA to the pathogen, whereby the pathogen-specific promoter directs transcription of the ribozyme(s) 20 in the cells of the pathogen.

A method of treating an infection in a subject is provided, comprising administering to the subject the liposome comprising DNA comprising a target-specific promoter and encoding a ribozyme, whereby the ribozyme encoded by the DNA is expressed and the infectious agent is killed or weakened. The liposome used in this method can comprise any 25 ribozyme-encoding nucleic acid, particularly those described herein targeted to genes of the pathogen. The infection can be bacterial, fungal, yeast, parasitic, viral or non-viral.

Parenteral administration, if used, is generally characterized by injection (intravenous, intradermal, subcutaneous and intramuscular). Injectables can be prepared in conventional forms, either as liquid solutions or suspensions, solid forms suitable for 30 solution of suspension in liquid prior to injection, or as emulsions. A more recently revised approach for parenteral administration involves use of a slow release or sustained release system such that a constant level of dosage is maintained. See, e.g., U.S. Patent No. 3,610,795, which is incorporated by reference herein.

Suitable carriers for parenteral administration of the substance in a sterile solution or 35 suspension can include sterile water or saline that can contain additives, such as ethyl oleate

or isopropyl myristate, and can be injected, for example, intravenously, as well as into subcutaneous or intramuscular tissues.

Topical administration can be by creams, gels, suppositories and the like. *Ex vivo* (extracorporeal) delivery can be as typically used in other contexts.

5 Oral administration is also contemplated. Suitable carriers for oral administration include one or more substances which can also act as flavoring agents, lubricants, suspending agents, or as protectants. Suitable solid carriers include calcium phosphate, calcium carbonate, magnesium stearate, sugars, starch, gelatin, cellulose, carboxypolymethylene, or cyclodextrins. Suitable liquid carriers can be water, pyrogen free
10 10 saline, pharmaceutically accepted oils, or a mixture of any of these. The liquid can also contain other suitable pharmaceutical additions such as buffers, preservatives, flavoring agents, viscosity or osmo-regulators, stabilizers or suspending agents. Examples of suitable liquid carriers include water with or without various additives, including carboxypolymethylene as a pH-regulated gel.

15 The multi-ribozyme can be administered to the subject in amounts sufficient to produce an antibiotic effect or to inhibit or reduce the activity of the target pathogen. Optimal dosages used will vary according to the individual, on the basis of age, size, weight, condition, etc, as well as the particular modulating effect being induced. One skilled in the art will realize that dosages are best optimized by the practicing physician and
20 20 methods determining dosage are described, for example, in Remington's Pharmaceutical Sciences (Martin, E.W. (ed.) Remington's Pharmaceutical Sciences, latest edition Mack Publishing Co., Easton, PA.). Treatment can be at intervals and can be continued for an indefinite period of time, as indicated by monitoring of the signs, symptoms and clinical parameters associated with a particular infection. The parameters associated with infection
25 25 are well known for many pathogens and can be routinely assessed during the course treatment.

6. EXAMPLES

30

Promoter Selection

Promoters specific for the targets are selected by screening genomic sequences as described, *supra*. For non-viral pathogens, plasmid capable of stable replication and maintenance in the microorganism was modified by standard molecular biology techniques
35 35 to carry the coding region of a reporter gene (Sambrook et al., *supra*). The reporter gene is any standard reporter gene known in the art including but not limited to the lacZ gene of *E.*

coli, which codes for β -galactosidase. Total genomic DNA is isolated from cells of the pathogen, cleaved with restriction endonucleases to yield fragments of a few hundred basepairs on average. These fragments are then ligated into a unique restriction endonuclease cleavage site at the 5' end of the reporter gene coding region, creating a 5 library of plasmids. The library is then transformed into the pathogen by standard techniques and the resulting transformants are screened for expression of the reporter gene. In the case of lacZ, the transformants can be plated onto medium containing the chromogenic-galactosidase substrate X-Gal (5-bromo-4-chloro -3-indolyl-D-galactoside). Transformants that contain a plasmid with an insert carrying a promoter will express β - 10 galactosidase and will turn blue on X-Gal plates. The intensity of the blue color is relative to the level of expression; promoters of different strength can be selected based on the 15 intensity of the blue color.

To test for species-specificity, the identified promoters are transferred to promoterless reporter plasmids capable of replication and maintenance in a different 15 organism. Truly species-specific promoters will not activate the expression of the reporter gene in any other species. Obvious modifications can be used to identify and test artificial promoters composed of synthetic oligonucleotides inserted into the promoterless reporter plasmid.

Several pathogen-specific promoters have been discovered by the inventors to be 20 useful in the methods and compositions of the invention and serve to illustrate the present invention. Three pseudomonas promoters include ARN, PROC, and ARC.

ARN: 5' ACTCGCGGA TCATCTTCAC CATCGGCCGC AACTCCTGCG
GGATATCCTC GTCCTCCTCC TCCACCGGCA CCCCCATGGT AGCGGCCAGC
25 TCGCGCCCTG CCTGGGAAAG CTGTACATGC TGATCGCGG CGTCGGTGCC
GGCGGCCGGG TCTTCCGCCT GCTCGCGGT GCCGGTCCGT GCGGCCTTGG
CGTCCGCGGC GGCGCGCGAT GAGGGCGGCA CCTGGGTGGT GATCCAGCCA
CTGAGGGTCA ACATTCCAGT CACTCCGGGA AAAATGGAAT TCTTCCATTG
GATCGGCCCA CGCGTCGCGA ACTTGAGCCC CCTTTCGTC GCCCCTTGAC
30 AGGGTGCAC AGGTAGTCGC AGTTGTTGA CGCAAGTCAC TGATTGGAAA
CGCCATCGGC CTGTCAGAAA TGGTCGTTGCC AGACCTATGG CTGGCACCCG
CATCGCGGCT GCGTTACCCCT TACTCCTGTT GTGCCTTAA CCTAGCAAGG AC
(SEQ ID NO:1)

PROC: 5' AATTCCCTCGA AGTCCTTGCCTGCTGTTGTCG TTCAATGATGT
CGTAGATCAG CGCATGCACC TGCTTGTGTT CCAGCGGTGG CAGGTTGATC
CGGCGTACAT CGCCATCCAC CCGGATCATG GGTGGCAGGC CGGCGGAGAG
GTGCAGGTCC GAAGCGCCCT GTTGGCACT GAAGGCGAGC AGCTCGGTAA
5 TATCCATGGG ACTCCCCAAT TACAAGCAAG CAGGTAGAAT GCCGCCAAAG
CCGCCGTCTC GGACAAGGAA AACACCGGAT GAGCCAGGGT GCTTCCAGGA
CACCGTGGT GTCCTGCGCC AGACGCGAA CCTCGACACT GGAACAGGAA
GATGGCCATC GAGGCCGGCG GTTTCGAGGG CGTCGAGCCG ACGCCGACCG
CACTTCCATA GGGCGCAGGT AATGTCCACG ATAGCAGAGA ATATTGCAAA
10 GGTTGCCGCG CGCATCCGTG AGGCAGCGCA AGCTGCCGGG CGCGATCCGG
CCACGGTCGG CCTGCTCGCC GTGAGCAAGA CCAAGCCGC CGCCGCCGGT
CGCGAGGCAGC ACGCCGCCGG CCTTCGCGAC TTCGGCGAAA ACTACCTGCA
GGAGGCCCTC GGCAAGCAGG CCGAACTGGC CGACCTGCCCG TTGAACCTGGC
ACTTCATCGG CCCCATCCAG TCGAACAAAGA CGCGGCCCAT CGCCGAGCAT
15 TTCCAGTGGG TGCACTCGGT GGACCGGTTG AAGATCGCGC AGCGCCTGTC
GGAGCAACGC CGGGCCGGGC TGCCGCCCT GAATGTCTGC CTGCAGGTCA
ACGTCAGCGG CGAACGCCAGC AAGTCCGGCT GCGCCCCCGA GGACCTGCCG
GCCCTGGCCG AGGCCGTGAA GCAACTGCC AACCTCCGAT TGCCTGGCCT
GATGGCCATC CCCGAACCCA CCGCCGAACG CGCCGCGCAA CACGCCCGT
20 TCGCCCGCCT GCGCGAACTG CTGCTGGACC TGAACCTTGG CCTGGACACC
CTGTCCATGG GCATGAGCGA CGACCTCGAG GCAGCCATCGG CGAAGGTGCG
ACCTGGGTCC GCATCGGTAC CGCCCTGTT GGCGCCCGCGA CTACGGCGCG
CCGGCTTCTT GAATGAATCCC
(SEQ ID NO:2)

25

ARC: 5' CTAGAGCTAT TGATGTGGAT CAACATTGTC CACTAGCCGC
TGCCGCCTAA TCTCCAGAAT TGTGAG
(SEQ ID NO:3)

30 The invention provides modifications of the ribozyme cassettes described herein. In
the following example, the pChop cassette has been modified to include a spacer insert
(~300 bases of DNA between the Eco and Bam sites) as well as a modified hairpin on the 3'
end of structure, and a TL17 terminator sequence. Without limitation, the spacer insert
facilitates cloning and preparation of the vector with decreased background. Generally, the
35 spacer insert is excised prior to cloning in an insert (such as a sequence from a library). The
modified hairpin provides protection against 3'->5' exonuclease activity. The hairpin loop

is liberated along with the trans-acting ribozyme(s) and forms a secondary protective structure. The TL17 terminator sequence serves to stop prokaryotic transcription following expression of the entire modified pChop cassette, and may act as a transcriptional terminator in both directions (thus preventing reverse read-through transcription). The 5 modified cassette, indicated as UPCM2, below, was linked to the LESHI promoter (see U.S. patent application serial no. 09/291,902, filed April 14, 1999, incorporated herein by reference in its entirety). The cassette was further cloned into pBluescript II sk vector (Statagene, Inc.) Further, an AP LacI element was cloned downstream of the cassette. The AP promoter is a strong constitutive promoter that linked to Lac I. Lac I expression 10 provides tighter regulation (e.g., in the uninduced state).

The modified cassette was used in *in vivo* ribozyme library screening. A hammerhead ribozyme library was constructed with degenerate bases for the two antisense arms (helix I and III), such that the library contains essentially all ribozymes capable of binds to and cleaving any stretch of accessible RNA sequence containing the NUX motif. 15 The library was constructed from a custom synthesized single stranded oligonucleotide. The ribozyme library represents approximately 6.7×10^7 unique hammerhead ribozyme sequences. A second strand was synthesized *in vitro* utilizing a compatible primer and polymerase. The double-stranded oligonucleotide was then trimmed with restriction endonucleases to allow for directional cloning into the plasmid vector, the modified pChop 20 cassette vector, prepared with compatible cohesive ends. Additionally, a lacI gene was cloned onto the plasmid vector to allow for tighter repression of lac operator containing modified pChop expression promoters. The ribozyme library was then ligated into the vector (modified pChop cassette on a pBluescript backbone plasmid) and this ligation is transformed into *E. coli* host cells. The cells were plated onto a dish containing solid media 25 with the Ampicillin antibiotic selection and allowed to grow until small, isolated colonies were present. The dish was then replica plated using sterile velvet onto solid media containing the inducer compound, IPTG. Replica plates are incubated for approximately equivalent time as the parent plate. Following incubation, parent and replica plates were compared, and putative positive colonies (those showing a difference between the parent 30 and replica plate) were picked and propagated from the original parent plate by standard techniques in the art. The highest ratio of induced versus repressed expression levels was in *E. coli* S03831 slow-ribosome cells. The sequence of the ribozyme of the picked colony was then determined by standard methods in the art. The RNA target was also determined. Once sequenced, ribozymes revealed in the screen were further redesigned and refined to 35 enhance the activity of the ribozymes.

UPCM2 cassette sequence

5'-TCAGAAAATTATTTAAATTCCAATTGACATTGTGAGCGGATAACAATATAA
TGTGTGGAAGCTTATCGATACCGTCGACCTCGAAGCTTGGAACCCCTGATGAGT
5 CCGTGAGGACGAAACGATGACATTCTGCTGACCAGATTACGGTCAGCAGAAT
GTCATCGTCGGTCCAGGATCCGGCTGCTAACAAAGCCGAAAGGAAGCTGAG
TTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAACCCCTGGGCCTCTAAA
CGGGTCTGAGGGGTTTTGCTGAAAGGAGGAACATATCCGGATATCCCGCA
AGAGGCCCGGCAGTACCGGCATAACCAAGCCTATGCCTACAGCATCCAGGGT
10 ACGGTGCCGAGGATGACGATGAGCGCATTGTTAGATTACATACACGGTGCCTGA
CTGCGTTAGCAATTAACTGTGATAAAACTACCGCATTAAAGCTTATCGATGATA
AGCTGTCAAACATGAGAATTGGCGTATACGCCGAATTCAAGGGTCTGCGCA
ACGACGACGATGAGGTACCACATCGTCGTTGCGCACTGATGAGGCCGTGA
GGCGAAACCCCTGACGCGTAAAAAAACCCGCCCCGGGGTTTTACCCCTT
15 CCTATGCGGCCGCTCTAGTCGAGGGGGGGCCGCTAGAACTAG-3'
(SEQ ID NO:4)

A second modified cassette that has been constructed is the P2CM2, indicated below, which provides a modified pChop cassette which has been modified to include a 20 spacer insert (~300 bases of DNA between the Eco and Bam sites) as well as a modified hairpin on the 3' end of structure, and a TL17 terminator sequence. The second modified cassette, indicated as UPCM2, below, was linked to the P2 promoter.

P2CM2 cassette sequence

25 5'-AGAAAGCAAAAATAATGCTTGACACTGTAGCGGAAGGCGTATAATGGAA
TTGTGAGCGGATAACAATTACAAGCTTATCGATACCGTCGACCTCGAGCTTG
GAACCCCTGATGAGTCCGTGAGGACGAAACGATGACATTCTGCTGACCAGATT
CACGGTCAGCAGAATGTCATCGTCGGTCCAGGATCCGGCTGCTAACAAAGC
CCGAAAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAAC
30 CCCTGGGCCTCTAAACGGTCTTGAGGGTTTGCTGAAAGGAGGAACTA
TATCCGGATATCCCGCAAGAGGCCGGCAGTACCGCATAACCAAGCCTATGC
CTACAGCATCCAGGGTACGGTGCCGAGGATGACGATGAGCGCATTGTTAGAT
TTCATACACGGTGCCTGACTGCGTTAGCAATTAACTGTGATAAAACTACCGCAT
TAAAGCTTATCGATGATAAGCTGTCAAACATGAGAATTGGCGTACGCCGA
35 ATTTCAAGGGTCTGCGCAACGACGACGATGAGGTACCACATCGTCGTTGCG

CACTGATGAGGCCGTGAGGCCGAAACCCTGACGCGTAAAAAAACCGCCCC
GGCGGGTTTTACGCGTTCCTATGCGGCCGCTCTAG -3'
(SEQ ID NO:5)

5 Example of Tissue-specific Multi-Ribozymes

In one embodiment of the present invention, a multi-ribozyme is engineered to down-regulate a targeted RNA in a tissue-specific manner. In accordance with this embodiment, the multi-ribozyme may be designed to target sebum production and secretion and be designed for topical administration, which will down regulate sebum production via 10 selective expression within the pilosebaceous unit. The DNA construct, placed within a suitable expression vector, comprises a tissue-specific promoter (from keratin K4, K7, or K13) driving expression of multi-ribozymes targeted against the androgen receptor (AR) and/or the enhancer protein steroidogenic factor 1 (SF1). In one embodiment of the invention, the triple ribozyme cassette consists of two cis-acting ribozymes flanking a trans- 15 acting internal ribozyme. The action of the cis-acting ribozymes liberates the internal targeted ribozyme with minimal flanking sequences, thereby enhancing its activity.

Sebum production and secretion is an important component in the pathogenesis of acne, and sebum production is regulated by androgens. Thus, the topical application of a formulation containing the multi-ribozyme of the present invention results in the inhibition 20 of excessive sebum production/secretion by sebaceous glands in skin.

The approach utilized is to create a DNA construct within a suitable expression vector, which comprises a tissue-specific promoter driving expression of triple ribozymes, or multi-ribozyme of the invention which are targeted to cellular components important in sebum production. One target is the androgen receptor (AR), which is known to be 25 important in the process. The Androgen Receptor is necessary for the androgen responsiveness and metabolism of androgen to activated forms, and metabolic processes which are controlled by enzymes whose expression is controlled by steroidogenic factor 1 (SF-1). The other target is steroidogenic factor 1 (SF1), which is an enhancer protein important for steroid metabolizing enzymes in the pathway which activates testosterone. 30 The promoters utilized are those which drive expression of keratins 4, 7, and 13 (K4, K7, and K13); expression of these keratins are quite selective for sebaceous glands within the pilosebaceous unit, given the distribution of topically applied liposomal formulations. In one example of the invention, the triple ribozymes consist of two cis-acting ribozymes flanking an internal transacting ribozyme, which is targeted to AR and/or SF1. The two cis- 35 acting ribozymes liberate the internal targeted ribozyme(s) with very short nonspecific flanking sequences, thereby enhancing its efficiency and specificity.

Biologic Delivery

The multi-ribozymes of the present invention may be delivered by a wide variety of viral vectors and bacteriophage as described herein. In one embodiment, the multi-ribozymes are delivered via a plasmid encoding the ribozymes, a plasmid origin of replication, a selectable marker for plasmid maintenance, the minimal lambda origin of replication, and cos sites, which are required for packaging of DNA into lambda virions. 5 This plasmid is maintained in a lambda lysogen that is defective in integration/excision and recombination functions. The defective lysogen provides all of the replication factors needed to activate the lambda origin of replication on the plasmid and all of the structural components needed to form mature virions; however, the lysogen is not able to replicate 10 and package its own DNA into the virions. The lysogen also carries the cl⁸⁵⁷ temperature-sensitive repressor mutation. Induction of the lysogen by temperature shift to 42°C or by other means, such as exposure to 5J/m² of ultraviolet radiation will mobilize the plasmid 15 and result in its replication and packaging into lambda virions. The virions are then harvested, purified free of *F. coli* proteins and are used to deliver the ribozyme gene(s) or nucleic acid to *E. coli*.

Abiologic Delivery

Abiologic delivery of the Multi-ribozyme is accomplished with ribozyme(s) 20 constructs that have been engineered to be expressed within the targeted tissue. Briefly, the genetic element containing the promoter and ribozyme(s) are complexed with cationic liposomes (Lipofectamine, Gibco BRL) in a 1:10 ratio and are introduced into test animals by either single or multiple injection of 0.2 ml total volume nucleic acid-liposome mixture.

25 *In Vivo* Testing

Following the demonstration that multi-ribozymes of the present invention have an *in vitro* biological activity (either directly on bacterial cultures or in an infectious tissue culture cell assay system), the effectiveness of the multi-ribozymes, is shown in an *in vivo* model system. To demonstrate the efficacy of multi-ribozyme *in vivo*, experimental animal 30 model systems are utilized. For an initial demonstration of the efficacy of the multi-ribozyme *in vivo*, mice are infected with a microbial pathogen which has previously been shown to be sensitive to the multi-ribozyme construct(s) and the effect of multi-ribozyme administered *in vivo* is determined. In the first series of *in vivo* trials, one determines the effectiveness of a multi-ribozyme at preventing an acute infection in a murine model system 35 when the multi-ribozyme is added directly to the microbe prior to administration *in vivo*. Examples of model systems that are useful in connection with the present invention include

but are not limited to those presented in Abdul-Hassan, et al., 1990, Bacteriophage therapy of pseudomonas burn wound sepsis. Annals of the Mediterranean Burn Club. 34:262-264; Kwarcinski, W., B. Lzarakiewicz, B. Weber -Dabinwoska, J. Rudnicki, K. Kaminski, and M. Sciebura. 1994, Bakteriofagoterapia w. leczeniu nawracajacego ropnia

5 podprzeponowego i podwątrobowego oraz przetoki jewlitowej Patent Office wycieciu zoladka. Polski Tygodnik Lekarski. XLIX:23-23;Slopek, S., B. et al., 1987, Results of bacteriophage treatment of suppurative bacterial infections in the years 1981-1986, Arch. Immunol. Ther. Exp. (Warsz) 35:569-583; Smith. H.W., and M.B. Huggins, 1982, Successful treatment of experimental Escherichia coli infections in mice using phage: its

10 general superiority over antibiotics. J. Gen. Microbiol. 128:307-318; and Soothill, J.S., 1994, Bacteriophage prevents destruction of skin grafts by Pseudomonas aeruginosa. Burns. 20:209-211.

The next series of trials determine whether the administration of multi-ribozyme

15 after infection is effective at preventing an acute bacterial infections. In addition to the clinical status of infected mice, tissues obtained at necropsy are examined histologically and the presence of replicating microorganism in tissue samples is determined by standard methodology. Animals can be infected by various routes (systemic and/or mucosal) and the multi-ribozyme delivered over time after infection by systemic and/or mucosal routes. Both

20 abiologic as well as biological delivery of multi-ribozyme is used. The demonstration of a positive effect of the multi-ribozyme in controlled experimental model system provides compelling evidence for the efficacy of the preparation and determines whether or not the preparation warrants evaluation under conditions of standard clinical trials.

25 Development and Testing of the Catalytic Component of the Multi-ribozyme

The following is a routine approach for designing, manufacturing and testing of the ribozymes that are incorporated into the multi-ribozyme invention.

The catalytic component of multi-ribozyme invention is/are trans-acting internal targeted ribozyme (ITRz). To facilitate construction of this critical and catalytic

30 component, the targeted triple ribozyme (TRz) containing a double ribozyme cassette was developed, as shown in Figure 1. This artificially contrived genetic element consists of autocatalytic, self-cleaving 5' and 3' ribozymes, with a cloning region (denoted by the box entitled Targeted Ribozyme) between them. This double ribozyme cassette was then placed within a series of expression vectors that were either constructed (pClip), or modified from

35 purchased from commercial vendors (pBluescriptII, Stratagene; pCRII, InVitrogen; pET-30a-c; pBACsurf-1, pIE1 and pIE4, Novagen) and used intact or modified as necessary to

confer the desired activity within the Multi-ribozyme. pClip (the genetic element described in Figure 2, was a modification of pBluescript, wherein the cassette shown is cloned into the Not I site in pBluescript. The targeted ribozyme (transacting catalytic ribozyme) was cloned into the Bgl II site (TGCTCT). Liberation of internal ribozymes from pClip resulted 5 in a distribution of the catalytic core ribozyme(s) to approximately 20% nuclear and 80% cytoplasmic.

A second ribozyme cassette that was constructed was pChop. pChop was modified from pClip to convey a more efficient and effective liberation of the internal transacting ribozymes. The pChop ribozyme cassette is diagramed in Figure 3. Liberation of internal 10 catalytic core ribozymes from pChop increased localization to the nucleus.

A third ribozyme cassette that was constructed was the pSnip ribozyme cassette. The pSnip multi-ribozyme was constructed by engineering the pClip cassette 5' to pChop. In addition, the pSnip multi-ribozyme contained a catalytic core sequences with two trans-acting ribozymes in each cassette. Each pair of trans-acting ribozymes was linked by 15 a short spacer and stabilized by a hairpin loop located 3' to the pair. Figure 4 diagrams the schematic of the pSnip cassette. Figure 5 diagrams the nucleotide sequence of the pSnip cassette indicating the sequence of the cis-acting cassette minus the internal ribozyme sequence.

An internal targeted ribozyme (ITRz) was synthesized as reverse complementary 20 overlapping oligodeoxynucleotides, which were designed in such a way that when annealed they form single stranded ends identical to those produced by digestion with the restriction endonuclease contained with the region between the two cis-acting ribozymes. In this particular example the restriction endonuclease recognition site was that recognized by Bgl II.

25 An important advantage of the present invention is that essentially any RNA can be targeted, since specificity is conferred by selecting sequences for the ribozyme that are reverse and complementary to sequences flanking the chosen cleavage site in the targeted RNA molecule. The internal targeted ribozymes are then cloned into the cloning region within the double ribozyme cassette to produce the targeted trans-acting ribozyme. Internal 30 targeted trans-acting ribozymes to prokaryotic sequences have been constructed including, but not limited to, *Escherichia coli*: *secA* (EcosecA, AE000119 U00096), gene X (EcosecA, AE000119 U00096) *ftsZ* (AE000119;U00096), *dnaG* (AE000388 U00096), *rpoA*(AE000407 U00096) and tRNA-asp (X14007), *Streptomyces lividins secA* (Z50195), *Enterococcus faecalis, ftsZ* (U94707) *Pseudomonas putida, dnaG* (U85774), *Streptomyces* 35 *coelicolor rpoA* (X92107), *Staphylococcus warneri tRNA, Asp* (X66089 S42075), *Staphioccocus RNA III*.

Ribozyme Activity and Liberation of Internal Ribozymes

As shown in the autoradiograph of Figure 7, the catalytic activity of ribozymes which are attached at either their 5' or 3'ends is not diminished. The constructs tested are diagramed in Figure 6, 50nM of the each of the constructs were incubated for 37 °C for 0.5 5 or 2 hours with target RNA (at about 50nM). In this example, the transacting ribozymes of the constructs were targeted to multi-catalytic proteinase component C9. Following incubation, samples were denatured and separated on a polyacrylamide gel. The upper band represents the uncleaved target RNA whereas the lower bands represent the cleaved products. The data demonstrate that a ribozyme attached at either the 5' (lane 2) or 3' (lane 10 4) end of a transacting ribozyme does not diminish the catalytic activity of the ribozyme.

RT/PCT Analysis of TRz Expression and Self-Liberation in B2-X Clones. Cells were stably transfected with a B2-targeted PCLIP multi-ribozyme. Figure 8A shows cytoplasmic RNA results, while Figure 8B shows nuclear RNA results. These results demonstrate that there was a distribution of liberated transacting ribozymes between the 15 nucleus and the cytoplasm. Most clones show 10 to 20% of the transacting ribozyme in the nucleus, although B2-1 was mostly cytoplasmic. 18s ribosomal RNA was amplified to document equivalent amounts of RNA.

An RT/PCR protocol was used for quantification of Rz expression, as well as for assessment of self-liberation of the ITRz *in vivo*. An "inner" set of primers was used in the 20 RT/PCR reactions; these primers lie internal to the self-cleavage sites, and they effectively amplify both processed and unprocessed TRz transcripts. The upstream primer is 5'-AGCTCGAGCTCAGA (SEQ ID NO:6), and the downstream primer is 5'-TCGACGGATCTAGATCC (SEQ ID NO:7). Following self-cleavage of TRz transcripts, these primers were non-functional, so that they detect only unprocessed 25 transcripts. To provide a baseline for the relative efficiency of the primer pairs, RT/PCR amplifications were performed with a mutant that does not undergo self-processing. One inner or outer primer was end labeled with ^{32}P , and RT/PCR amplifications were performed with 2 μg cytoplasmic RNA. Following the reactions, samples were separated by PAGE in 6% polyacrylamide gels, the gels were dried and the radioactivity was detected by 30 autoradiography and also using a phosphorimager. Product obtained with the inner primer pair was 1.8X greater than that obtained with the outer primer pair.

The following sequences are for ribozymes directed against the targets described. The naming system refers to the target cytosine in the GUC motif. It is the nucleotides 35 number from the referenced sequence (accession number indicated). Ribozymes directed against *secA* targets have restriction sites for Bgl II on both ends. All other inserts have Bgl

II (5' end) and Sty I (3' end) restriction sites for use in the new vector. Antisense arms are boldfaced.

Escherichia coli

5 ***ftsZ* target** (ACCESSION: AE000119 U00096)
105
AGATCTAAACGCCGATCTGATGAGTCCGTGAGGACGAAACTTAAAAACCAAGG
713
AGATCTAAACATCTCACTGATGAGTCCGTGAGGACGAAACATTACGAAACCAAAAGG
1131
10 AGATCTAAATCATTCACCTGATGAGTCCGTGAGGACGAAACTTTAGCAAACCAAGG
(SEQ ID NO:8)

secA target (ACCESSION: AE000119 U00096)
84
15 AGATCTAAAAAAAACCTGATGAGTCCGTGAGGACGAAACTGGTAAAAGATCT
707
AGATCTAAATTATCCACTGATGAGTCCGTGAGGACGAAACGGCGAAAAGATCT
856
AGATCTAAATCGTTACCTGATGAGTCCGTGAGGACGAAACTACCGAAAAGATCT
20 894
AFATCTAAATGATGTTCTGATGAGTCCGTGAGGACGAAACCACTTAAAAGATCT
979
AGATCTAAATTCCACTGATGAGTCCGTGAGGACGAAACGTGCAAAAGATCT
1282
AGATCTAATTGATACCCTGATGAGTCCGTGAGGACGAAACAGTCAGAAAAGATCT
25 2216
AGATCTAAATTCGTTCTGATGAGTCCGTGAGGACGAAACACCACAAAAGATCT
(SEQ ID NO:9)

dnaG target (ACCESSION: AE000388 U00096)
30 5344
AGATCTAAACGTTAGTCTGATGAGTCCGTGAGGACGAAACCAACAAAACCAAGG
5903
AGATCTAAAGGCATCACTGATGAGTCCGTGAGGACGAAACTGTTAAAACCAAGG
6336
AGATCTAAACCACATCCTGATGAGTCCGTGAGGACGAAACAGTTAAACCAAGG
35 (SEQ ID NO:10)

rpoA target (ACCESSION: AE000407 U00096)

8308

AGATCTAAAAGAGCGCTGATGAGTCCGTGAGGACGAAACAGTCAAAACCAAGG

8494

5 AGATCTAAATTGATCTGATGAGTCCGTGAGGACGAAACAGCTAAACCAAGG

8737

AGATCTAAACGATTCTGATGAGTCCGTGAGGACGAAACATCACCAAACC AAGG

(SEQ ID NO:11)

10

tRNA-Asp target

(directed against GUC anticodon loop. Accession: X14007)

172

AGATCTAAATGCGTCTGATGAGTCCGTGAGGACGAAACAGGCAGGTAAAACCAAGG

15 (SEQ ID NO:12)

Streptomyces lividans

secA target (ACCESSION: Z50195)

1080

20 AGATCTAAACTCGTCCTGATGAGTCCGTGAGGACGAAACGATCAAAACCAAGG

2033

AGATCTAAAGGGCGCTGATGAGTCCGTGAGGACGAAACGCGAAAACCAAGG

2556

AGATCTAAAGTACTCCTGATGAGTCCGTGAGGACGAAACCGAGCGAAACCAAGG

25 (SEQ ID NO:13)

Enterococcus faecalis

ftsZ target (ACCESSION: U94707)

30 10805

AGATCTAAAACATAATGCTGATGAGTCCGTGAGGACGAAACGAGTTAAAACCAAGG

11182

AGATCTAAAGTTAATAACTGATGAGTCCGTGAGGACGAAACTTGTCAAACCAAGG

11512

AGATCTAAAACTTTGCTGATGAGTCCGTGAGGACGAAACGTGTATAACCAAGG

35 (SEQ ID NO:14)

Pseudomonas putida

dnaG target (ACCESSION: U85774)

222

5 AGATCTAAAGGTCCATCTGATGAGTCCGTGAGGACGAAACAAAGCAAACCAAGG

986

AGATCTAAACAGGTTCCCTGATGAGTCCGTGAGGACGAAACAATGTAAACCAAGG

1891

AGATCTAAATCGCTTCTGATGAGTCCGTGAGGACGAAACGTGATAAACCAAGG

(SEQ ID NO:15)

10

Streptomyces coelicolor

rpoA target (ACCESSION: X92107)

290

15 AGATCTAAAGCTCGATCTGATGAGTCCGTGAGGACGAAACGAACCAAACCAAGG

716

AGATCTAAACGAGTCCTGATGAGTCCGTGAGGACGAAACCGGGAAACCAAGG

1099

AGATCTAAAGTCGATGCTGATGAGTCCGTGAGGACGAAACTTCGCAAACCAAGG

(SEQ ID NO:16)

20

Staphylococcus warneri

tRNA-Asp target

(directed against GUC anticodon loop. Accession: x66089 s42075)

25 62

AGATCTAAATGCGTCTGATGAGTCCGTGAGGACGAAACAGGCAGGCGAAACCAAGG

(SEQ ID NO:17)

The utility of the design using eukaryotic sequences has also been evaluated in the
30 following examples: a) repetitive B2 transcripts (B2); b) RNA polymerase I (polI); c)
Hepatitis B virus (HBV); d) Sonic Hedgehog (SH); e) Human Papillomavirus E6/E7 protein
(HPV); f) RNA polymerase II (polII); g) Insulin-like Growth Factor 1 (IGF1); h)
retinoblastoma protein (RB); i) and j) Multicatalytic Proteinase alpha-subunits C3 and C9
(C3 and C9, respectively); k) telomerase (tel); l) Transforming growth factor beta (TGF β);
35 m) catalase (CAT); n) Peroxisome proliferation associated receptor (PpaR α); and o)
Cytochrome P₄₅₀ 1E1 (p4501E1). Target RNAs (with locus names and accession numbers)

as well as the selected target sites are presented (Table 1), as are the sequences of these ITRz (SEQ ID NOS:18-36).

TABLE 1. Summary of Targeted RNAs and Target Sites.

5	Target RNA	EMBL Locus	Accession	Target Site	Functional Testing	
					<i>in vitro</i>	<i>in vivo</i>
10	pol II	HSRNAP14K	Z27113	GTC ₈₃	ND	ND
	HBV	XXHEPAV	X02496	GTC ₄₃₈	IP	+
15	RB	MUSP105RB	M26391	GTC ₂₆₄	+	+
	IGF1	HUMIGF1B	M37484	GTC ₁₈₅	ND	ND
	SH	MMEVX1	X54239	GTC ₅₅₈	IP	IP
20	Pol I	MUSRPA40	D31966	GTC ₄₅₈	+	+
	HPV	PPH16	K02718	GTT ₁₀₈	IP	+
	C3	RATC3AA	J02897	GTT ₂₂	+	+
	C9	RNPTSC9	X533304	GTC ₁₀₁	+	+
	B2	B2 -Consensus	##	GTT ₂₄	+	+
	Tel	MMU33831	U33831	CTA ₆₃	ND	ND

see Clawson, G. et al. Cell Growth Diff. 7:635-646 (1996).

Multiple target sites have been selected for TGFb, CAT, PpaRa, and p4501E1. All of these ribozymes (with the exception of TGFb) have been functionally tested *in vitro*. Ribozymes targeted to catalase have also been tested *in vivo*. *in vitro* testing refers to target cutting. *in vivo* testing refers to cell culture experiments or transgenic animals (for polI). IP, in progress. ND, not yet determined. +, substantially decreased target RNA (and/or protein).

B2 TGCTCTT CTGATGAGTCCGTGAGGACGAAA CCGCCTGA
(SEQ ID NO:18)

30 Pol 1 TTCAAAGA CTGATGAGTCCGTGAGGACGAAA CGAGGATC
(SEQ ID NO:19)

Sonic Hedgehog GTCCCAT CTGATGAGTCCGTGAGGACGAAA CCGGC
(SEQ ID NO:20)

	HBV	ATTAGAG CTGATGAGTCCGTGAGGACGAAA CAAACG (SEQ ID NO:21)
5	HPV	GTCCTGA CTGATGAGTCCGTGAGGACGAAA CATTGCA (SEQ ID NO:22)
	Pol III	TCCGTTGTCT CTGATGAGTCCGTGAGGACGAAA CATGACACCGA (SEQ ID NO:23)
10	IGF-1	GCGAGGAG CTGATGAGTCCGTGAGGACGAAA CATGGTGT (SEQ ID NO:24)
15	RB	AACTTT CTGATGAGTCCGTGAGGACGAAA CATAATG (SEQ ID NO:25)
C3	TCGAAGCTGT CTGATGAGTCCGTGAGGACGAAA CCGCGTTGA (SEQ ID NO:26)	
20	TEL	ATCAGGGT CTGATGAGTCCGTGAGGACGAAA GGTGCC (SEQ ID NO:27)
	C9	TCTTCGA CTGATGAGTCCGTGAGGACGAAA CATGGCT (SEQ ID NO:28)
25	TGF β -1	TAGCACA CTGATGAGTCCGTGAGGACGAAA CGTTGA (SEQ ID NO:29)
30	CAT/#13	TGCAATA CTGATGAGTCCGTGAGGACGAAA CTGCCT (SEQ ID NO:30)
CAT/#15	AAGTCAT CTGATGAGTCCGTGAGGACGAAA CCTGGA (SEQ ID NO:31)	
35	PpaRa/#2	GATAAGG CTGATGAGTCCGTGAGGACGAAA CTTTCC (SEQ ID NO:32)

PpaRa/#8 CATATTCTGATGAGTCCGTGAGGACGAAA CACTCG
(SEQ ID NO:33)

5 PpaRa/#14 TCATGTAT CTGATGAGTCCGTGAGGACGAAA CAAAAGG
(SEQ ID NO:34)

p4501E1/#2 GGTAAA CTGATGAGTCCGTGAGGACGAAA CTTGGG
(SEQ ID NO:35)

10 p4501E1/#8 GTCCAGT CTGATGAGTCCGTGAGGACGAAA CTTAAG
(SEQ ID NO:36)

For many of these constructs, "mutants" have also been created by substituting an A
15 for a G, or a G for an A, at nucleotides which are absolutely required for catalytic activity.
These "mutants" allow evidence indicating that the efficiency of destruction of the targeted
RNAs is due to ribozyme catalytic activity and not to antisense effects.

The eukaryotic cell contains large families of short repetitive sequences throughout
its genome.

20 Although B2 RNA transcripts are normally confined to the nucleus, they appear in
the cytoplasm of malignant cells, and in cell lines immortalized with SV40 or
papillomavirus as well as in undifferentiated embryonal carcinoma cells.

Multi-Ribozyme-Mediated Down Regulation of B2 Transcripts.

25 The B2 family represents a group of short repetitive sequences which are found
throughout the rodent genome and are analogous to the human Alu sequences. Certain B2
subfamilies are transcribed by RNA polymerase III, and this transcription is in part
controlled by the retinoblastoma protein. These actively transcribed B2 RNAs show a
predicted highly stable secondary structure.

30 The retinoblastoma protein gene protein (pRb) has recently been shown to act as a
regulator of pol III transcription 47, and pRb-mediated growth arrest has been hypothesized to
result from transcriptional blockade of tRNA synthesis, with consequent decreased protein
synthetic capacity. Therefore, it was important to determine whether the alterations
observed in B2 levels, by the inventors, might reflect alterations in pRb expression and/or
35 interactions.

Construction and *In Vitro* Testing of the B2-targeted Triple Ribozyme

For construction of the B2-targeted triple ribozyme, oligodeoxynucleotides were synthesized. The following oligodeoxynucleotides were used to construct the parent double ribozyme by the polymerase chain reaction;

5 primer 1: 5'-CCCGGGATTCTGATGGCCACGCCGGCCGCTCGA
GCTCTGATGAGTCCGTGAGGA-3' (SEQ ID NO:37)

primer 2: 5'-GACGGGATCCAGATCTGAGCTCGAGCTGACGGTA
CCGGGTACCGTTCTCGCCTCACGGA-3' (SEQ ID NO:38)

primer 3: 5'-GAGCTCAGATCTGGATCCGTCGACGGATCTAGATC
10 CGTCCTGATGAGTCCGTGAG-3' (SEQ ID NO:39)

primer 4: 5'-TTGCTTGGCCAGCGGCCGCTGCAGATCCGTTCTG
CCTCACGGACT-3' (SEQ ID NO:40)

Primer pairs 1 and 2, or 3 and 4 were annealed and amplified by PCR using a MiniCycler (MJ Research). PCR reaction products were then digested with *Bg*II and ligated with T4 15 DNA ligase. The resultant 173 bp fragment was isolated from a 3.75% low-melting temperature agarose gel (SeaPlaque, from FMC BioProducts) and subsequently purified using Wizard PCR kits (Promega). The recovered fragment was amplified by PCR using primer 1 and 4, and the PCR product was then digested with *Not*I and ligated into 100 ng 20 *Not*I-digested pOPRSVICAT mammalian expression vector (LacSwitch, from Stratagene), which contains a neomycin resistance gene. This parent double ribozyme was designated pClip.

The B2 targeted internal ribozyme was then made by annealing 100 pmol each of the primers:

5'-GATCTGCTCTGATGAGTCCGTGAGGACGAAACCGCTGA-3'
25 (SEQ ID NO:41) and
5'-GATCTCAGCGGTTCTCGCCTCACGGACTCATCAG AAGAGCA-3'
(SEQ ID NO:42). For annealing, primers were boiled for 3 minutes in 10 mM Tris-HCl, pH 8.5, 100 mM MgCl₂ and then slowly cooled to room temperature. After annealing, product was precipitated with ethanol and ligated into *Bg*II digested pClip at a 1:5 molar ratio. The 30 final pClip-B2 construct was verified by sequencing. For in vitro expression, the B2-targeted TRz was inserted into the *Not*I site of the pCRII vector, and M13 forward and reverse primers were used in PCR to amplify the region containing the B2-targeted Trz.

Characterization and Validation of the TRz construct

35 Typically, once the recombinant plasmid has been created the TRz constructs are isolated from the bacterium their nucleotide sequence is determined to confirm their

identities and to document their orientation within the vector. The constructs are then transcribed *in vitro* using SP6 and T7 RNA polymerases with ³²P-CTP. When transcribed in the "sense" orientation, all of these TRz constructs should be "self-liberating"; that is, the 5' and 3' self-cleaving autocatalytic ribozymes work effectively, freeing the ITRz during (or 5 immediately after) transcription (Figure 1c). The 5' liberated ribozyme (whose only function is self-cleavage, liberating the 5' end of the ITRz) is associated with relatively short stretches of vector sequences and the 3' self-cleaving ribozyme (whose only function is self-cleavage, liberating the 3' end of the ITRz) remains associated with long vector sequences. The liberated ITRz achieves its catalytic topology upon hybridization with the targeted 10 sequence. Transcription of all of these TRz in the "antisense" direction should not result in self-cleavage.

***In Vitro* Evaluation**

Upon validation of the TRz construct, the self-liberating TRz is evaluated for their 15 ability to effectively cut their targeted RNAs. Appropriate regions of the targeted RNAs are generally cloned using cellular RNA and reverse transcription/polymerase chain reaction amplifications. In some cases, cloned full-length cellular RNAs are also used. The identities of the constructs used for transcription of target RNAs are also confirmed by sequencing. Target RNAs are then synthesized *in vitro* using the appropriate T7/SP6 RNA 20 polymerase with ³²P-CTP, and are subsequently gel-purified. A preparation of the TRz under evaluation is then synthesized without ³²P-CTP. The TRz preparation is then mixed with their an appropriate concentration of radiometrically ³²P-labeled target or substrate RNA (³²P-labeled target RNAs and unlabeled TRz preparations are mixed at a 10:1 molar ratio) and is incubated for various lengths of time. Following incubations, the RNA is 25 examined using polyacrylamide gel electrophoresis (PAGE) and autoradiography. All of the constructs tested should be able to cleave their target RNAs. In general, the data show an approximate catalytic rate of 0.2 cleavages/ribozyme minute.

***In Vivo* Evaluation**

30 The TRz is evaluated with intact cells. The TRz cassette is excised from the parental plasmid and is then placed into an appropriate expression vector. Vectors utilized include (but are not limited to) the LacSwitch vector (from Stratagene), which is an IPTG-inducible system, and the TetSplice vector (from Gibco-BRL), which is a tetracycline-inducible system.

35 The TRz constructs in these expression vectors were then transfected into cells using standard techniques. Cell types used in transfections have included *E. coli*, human CaSki

cervical carcinoma cells, SV-40 immortalized rat hepatocytes, and mouse fibroblasts. In transient transfection analyses, all constructs tested produced substantial reductions in their respective target RNAs, thus demonstrating the effectiveness of the multi-ribozymes of the invention.

5

TRz construct 1:

The *secA* targeted TRz construct against the *secA* gene of *E. coli* is in the vector pClip, which is a variation of the generalized cloning vector, pBluescript of Stratagene. The plasmid containing the construct was transformed into competent bacterial cells and cells 10 containing the plasmid with the TRz were selected by using the antibiotic selectable marker within the vector, pClip. Upon induction of the promoter with isopropyl- β -D thiogalactoside, (IPTG) the effect of ribozyme expression is monitored by standard bacterial viable counts. A reduction in total viable cells is an indication of synthesis and catalytic activity of the TRz against the essential target.

15

TRz construct 2:

The poll-targeted TRz construct (in LacSwitch vector) was used in transfections of SV40-immortalized rat hepatocytes (CWSV1 cells), and stably transfected cell populations were obtained. Cells not transfected with the antibiotic resistance plasmid were all dead by 20 day 5, indicating that the antibiotic selection procedure was effective. Growth of cells transfected with the double ribozyme (as a control, with no cellular RNA target), and of cells expressing the catalytically inactive "mutant" poll TRz, was unaffected. However, growth of cells expressing the poll-targeted TRz was depressed by nearly 90%. Concurrently, the mRNA for poll was decreased by at least 70% from poll mRNA levels in 25 the cells expressing the mutant poll TRz. Since expression of the TRz was essentially equivalent for the two TRz, this clearly documents that the effects on cell growth are due to TRz catalytic activity and not to antisense effects. In other experiments, expression of the poll-TRz in LacSwitch resulted in cell death with mouse fibroblast cell populations.

30 TRz constructs 3:

The RB-targeted TRz (in the tetracycline-inducible TetSplice vector system) was used in transfections with CWSV1 cells, and stably transfected cells were selected using G418 antibiotic in the presence of tetracycline, and individual clones were harvested and used (expression of RB-targeted TRz is "off" in the presence of tetracycline, and is "on" in 35 the absence of tetracycline). Expression of the RB-targeted TRz had no effect on cell growth, as expected. Expression of RB mRNA was substantially reduced to below

detectable levels by Northern blot analysis. To extend this result, metabolic labeling of proteins was performed using ³⁵S-methionine, and immunoprecipitations were performed using an antibody directed against RB. The data show that RB protein levels were reduced by 75%, comparing clonal cell populations grown in the presence vs. absence of 5 tetracycline, even after only 18h of induction of RB-targeted TRz expression (removal of tetracycline). This shows that the effects of the reduction in RB RNA levels also extends to the production of the RB protein. In addition, the (+)-tetracycline sample provides an ideal control, since it represents the exact same clonal cell line. Essentially any inducible vector system can be used in parallel fashion.

10

TRz constructs 4:

The B2-targeted TRz (in the IPTG-inducible LacSwitch vector) was used in transfections of CWSV1 cells, and antibiotic selection was used to obtain a number of individual clones. Reductions in cytoplasmic B2 RNA levels of up to 80% were observed 15 by Northern blot analysis, and growth of transfected clones was reduced in parallel. In fact, a linear relationship between growth rate and B2 RNA levels was observed. The reductions in B2 transcripts paralleled the level of B2-targeted TRz expression (as determined by slot-blot analysis). The B2 target RNA is of additional interest, because B2 transcripts are not translated (i.e., they are not mRNAs) and they are abundant, highly-structured RNAs.

20 Other TRz constructs have also been successfully tested using this methodology (including C9 in the tetracycline-inducible system, and CAT, BRAC1 and Albumin driven by the albumin promoter in HepG2 cells).

Multi-Ribozyme-Mediated Down Regulation of NfkB p50 subunit target

25 Another target of the ribozymes of the invention is the NfkB p50 subunit. Six ribozymes (named RZ615, RZ636, RZ649, RZ1024, RZ1380 and RZ1438) were designed against the NfkB p50 subunit target. Results of *in vitro* cleavage assays, show in Figure 11, demonstrate that the ribozymes cleave the target NfkB p50 substrate, as indicated by the presence of fragments generated by following incubation of the ribozyme and substrate.

30 The *in vitro* substrate cleavage assay was performed by *in vitro* transcription of NFkB p50 subunit in the presence of P³² nucleotide, so the p50 transcript (substrate) was radiolabeled. The ribozyme was also transcribed *in vitro*. The ribozyme was then incubated with the P³²-labeled p50 messenger RNA for 45 minutes at 37°C. The reaction mixtures were then resolved by a 6% TBE-urea gel and exposed to film by methods known in the art.

35

The following sequences were constructed for the ribozyme constructs; the capitalized sequences correspond to the two arms of each ribozyme which match the target p50 sequence.

5 **RZ615:** 5' ctt gga acc gga tgc cag gca tcc ggt tGG TGC CTt tcg tcc tca
cg^g act cat cag TAG TGA a
(SEQ ID NO:43)

10 **RZ636:** 5' ctt gga acc gga tgc cag gca tcc ggt tAA GAA GTt tcg tcc tca
cg^g act cat cag TTA CCC Ta
(SEQ ID NO:44)

15 **RZ649:** 5' aat tca acc gga tgc cag gca tcc ggt tCT CAG GTt tcg tcc tca
cg^g act cat cag AAA ATC Tg
(SEQ ID NO:45)

20 **RZ1024:** 5' aat tca acc gga tgc cag gca tcc ggt tTG GAC CTt tcg tcc tca
cg^g act cat cag AGC GTG g
(SEQ ID NO:46)

RZ1380: 5' aat tca acc gga tgc cag gca tcc ggt tCA GCC Ttt cgt cct cac
gga ctc atc agT GTG TTg
(SEQ ID NO:47)

25 **RZ1438:** 5' aat tca acc gga tgc cag gca tcc ggt tAA CCT TTt tcg tcc tca
cg^g act cat cag CTC TAC g
(SEQ ID NO:48)

30 In order to demonstrate that the ribozymes designed against NF κ B were able to cleave the NF κ B mRNA and cause a decrease in NF κ B expression, the effect of Ribozyme-1024 (RZ1024) on the TNF α -induced production of IL-6 was assayed in A549 cells. TNF α is known to act via a signal transduction cascade to activate NF κ B. The activated NF κ B induces the expression of cytokines such as IL-6. Thus, a decrease in the production of IL-6 can act as a readout for one of the downstream effects of the NF κ B activation pathway.

A549 cells are human lung cancer cells, which express NF κ B and respond to NF κ B activation stimulus such as TNF α , LPS, etc. Ribozyme RZ1024 was cloned into the pCDNA 3.1 vector such that the expression of the ribozyme was driven by the CMV promoter. A549 cells were then stably transfected with the ribozyme RZ1024-pCDNA 3.1 construct. As depicted in figure 12, eight independent clones were established and designated as C2, C6, C7, C12, C19, C20, C21, and C22. The stable clones were then treated with TNF α (100ng/ml) for 7 hrs. Control (ctrl) represents untransfected A549 cells. After the TNF α treatment, media were collected for measuring secreted IL-6 using ELISA, and cells were harvested for total protein assay. As shown in figure 12, the TNF α -induced production of secreted IL-6 decreased, by 50% to 90%, in the eight clones transfected with ribozyme RZ1024 construct. Thus, the ribozymes of the invention are capable of targeting and cleaving a target RNA *in vitro* and *in vivo*.

Throughout this application various publications are referenced; the disclosures of the publications cited herein above are hereby incorporated by reference in their entireties into this application in order to more fully describe the state of the art to which this invention pertains.

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